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(71) Applicant: **PIONEER HI-BRED INTERNATIONAL, INC.** [US/US]; 800 Capital Square, 400 Locust Street, Des Moines, IA 50309 (US).

(72) Inventors: **FLANNAGAN, Ronald, D.**; 512 N.W. Norton Circle, Grimes, IA 50111 (US). **MATHIS, John, P.**; 3808 6th Street, Apt. 15, Des Moines, IA 50313 (US). **MEYER, Terry, EuClaire**; 4338 - 101st Street, Urbandale, IA 50322 (US).

(74) Agents: **SPRUILL, Murray, W. et al.**; Alston & Bird LLP, P.O. Drawer 34009, Charlotte, NC 28234-4009 (US).

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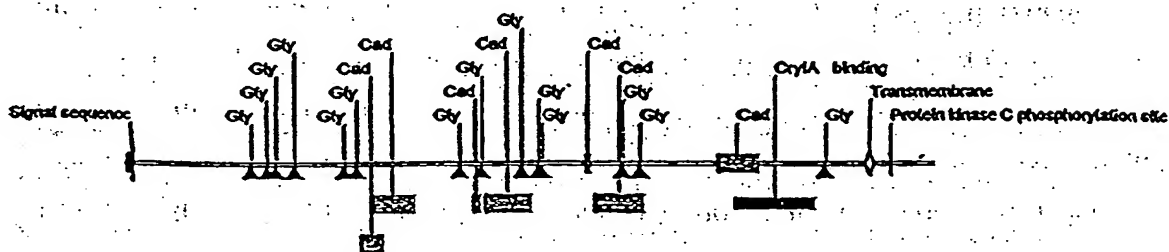
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(54) Title: **NOVEL BT TOXIN RECEPTORS FROM LEPIDOPTERAN INSECTS AND METHODS OF USE**



(57) Abstract: The invention relates to *Bt* toxin resistance management. The invention particularly relates to the isolation and characterization of nucleic acid and polypeptides for a novel *Bt* toxin receptor. The nucleic acid and polypeptides are useful in identifying and designing novel *Bt* toxin receptor ligands including novel insecticidal toxins.

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## NOVEL *Bt* TOXIN RECEPTORS FROM LEPIDOPTERAN INSECTS AND METHODS OF USE

### FIELD OF THE INVENTION

The field of the invention is manipulating *Bt* toxin susceptibility in plant pests. The field of the invention relates to the isolation and characterization of nucleic acid and polypeptides for a novel *Bt* toxin receptor. The nucleic acid and polypeptides are useful in developing new insecticides.

5

### BACKGROUND OF THE INVENTION

Traditionally, growers used chemical pesticides as a means to control agronomically important pests. The introduction of transgenic plants carrying the delta-endotoxin from *Bacillus thuringiensis* (*Bt*) afforded a non-chemical method of control. *Bt* toxins have traditionally been categorized by their specific toxicity towards specific insect categories. For example, the CryI group of toxins are toxic to Lepidoptera. The CryI group includes, but is not limited to, CryIA(a), CryIA(b) and CryIA(c). See Hofte *et al* (1989) *Microbiol Rev* 53: 242-255.

Lepidopteran insects cause considerable damage to maize crops throughout North America and the world. One of the leading pests is *Ostrinia nubilalis*, commonly called the European Corn Borer (ECB). Genes encoding the crystal proteins CryIA(b) and CryIA(c) from *Bt* have been introduced into maize as a means of ECB control. These transgenic maize hybrids have been effective in control of ECB. However, developed resistance to *Bt* toxins presents a challenge in pest control. See McGaughey *et al.* (1998) *Nature Biotechnology* 16: 144-146; Estruch *et al.* (1997) *Nature Biotechnology* 15:137-141; Roush *et al.* (1997) *Nature Biotechnology* 15 816-817; and Hofte *et al* (1989) *Microbiol Rev* 53: 242-255.

The primary site of action of CryI toxins is in the brush border membranes of the midgut epithelia of susceptible insect larvae such as lepidopteran insects. CryIA toxin binding polypeptides have been characterized from a variety of *Lepidopteran* species. A CryIA(c) binding polypeptide with homology to an aminopeptidase N has been reported from *Manduca sexta*, *Lymantria dispar*, *Helicoverpa zea* and *Heliothis virescens*. See Knight *et al* (1994) *Mol Micro* 11: 429-436; Lee *et al.* (1996) *Appl*

*Environ Micro* 63: 2845-2849; Gill *et al.* (1995) *J Biol. Chem* 270: 27277-27282; and Garczynski *et al.* (1991) *Appl Environ Microbiol* 10: 2816-2820.

Another *Bt* toxin binding polypeptide (BTR1) cloned from *M. sexta* has homology to the cadherin polypeptide superfamily and binds CryIA(a), CryIA(b) and CryIA(c). See Vadlamudi *et al.* (1995) *J Biol Chem* 270(10):5490-4, Keeton *et al.* (1998) *Appl Environ Microbiol* 64(6):2158-2165; Keeton *et al.* (1997) *Appl Environ Microbiol* 63(9):3419-3425 and U.S. Patent Patent No: 5,693,491.

A subsequently cloned homologue to BTR1 demonstrated binding to CryIA(a) from *Bombyx mori* as described in Ihara *et al.* (1998) *Comparative Biochemistry and Physiology, Part B* 120:197-204 and Nagamatsu *et al.* (1998) *Biosci. Biotechnol. Biochem.* 62(4):727-734.

Identification of the plant pest binding polypeptides for *Bt* toxins are useful for investigating *Bt* toxin-*Bt* toxin receptor interactions, selecting and designing improved toxins, developing novel insecticides, and new *Bt* toxin resistance management strategies.

## SUMMARY OF THE INVENTION

Compositions and methods for modulating susceptibility of a cell to *Bt* toxins are provided. The compositions include *Bt* toxin receptor polypeptides, and fragments and variants thereof, from the lepidopteran insects European corn borer(ECB, *Ostrinia nubilalis*), corn earworm (CEW, *Heliothis Zea*), and fall armyworm (FAW, *Spodoptera frugiperda*). The polypeptides bind CryIA toxins, more particularly CryIA(b). Nucleic acids encoding the polypeptides, antibodies specific to the polypeptides, as well as nucleic acid constructs for expressing the polypeptides in cells of interest are also provided.

The methods are useful for investigating the structure-function relationships of *Bt* toxin receptors; investigating the toxin-receptor interactions; elucidating the mode of action of *Bt* toxins; screening and identifying novel *Bt* toxin receptor ligands including novel insecticidal toxins; and designing and developing novel *Bt* toxin receptor ligands.

The methods are useful for managing *Bt* toxin resistance in plant pests, and protecting plants against damage by plant pests.

## BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 schematically depicts the location of the signal sequence, putative glycosylation sites, cadherin-like domains, transmembrane segment, CryIA binding region and protein kinase C phosphorylation site of the *Bt* toxin receptor from *Ostrinia nubilalis*; the  
5 nucleotide sequence of the receptor set forth in SEQ ID NO:1 and the corresponding deduced amino acid sequence in SEQ ID NO:2.

## DETAILED DESCRIPTION OF THE INVENTION

10 The invention is directed to novel receptor polypeptides that bind *Bt* toxin, the receptor being derived from the order *lepidoptera*. The receptors of the invention include those receptor polypeptides that bind *Bt* toxin and are derived from the *lepidopteran* superfamily *Pyraloidea* and particularly from the species *Ostrinia*, specifically *Ostrinia nubilalis*; those derived from *Spodoptera frugiperda* (*S.*  
15 *frugiperda*); and those derived from *Heliothus Zea* (*H. Zea*). The polypeptides have homology to members of the cadherin superfamily of proteins.

Accordingly, compositions of the invention include isolated polypeptides that are involved in *Bt* toxin binding. In particular, the present invention provides for isolated nucleic acid molecules comprising nucleotide sequences encoding the amino  
20 acid sequences shown in SEQ ID NOs: 2, 4, and 6; or the nucleotide sequences having the DNA sequences deposited in a plasmid in a bacterial host as Patent Deposit No. PTA-278, PTA-1760, and PTA-2222. Further provided are polypeptides having an amino acid sequence encoded by a nucleic acid molecule described herein, for example those set forth in SEQ ID NOs: 1, 3, and 5; those deposited in a plasmid  
25 in a bacterial host as Patent Deposit Nos. PTA-278, PTA-1760, and PTA-2222; and fragments and variants thereof.

Plasmids containing the nucleotide sequences of the invention were deposited with the Patent Depository of the American Type Culture Collection (ATCC), Manassas, Virginia on June 25, 1999; April 25, 2000; and July 11, 2000; and assigned  
30 Patent Deposit Nos. PTA-278, PTA-1760, and PTA-2222. These deposits will be maintained under the terms of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure. These deposits



were made merely as a convenience for those of skill in the art and are not an admission that a deposit is required under 35 U.S.C. §112.

The term "nucleic acid" refers to all forms of DNA such as cDNA or genomic DNA and RNA such as mRNA, as well as analogs of the DNA or RNA generated using nucleotide analogs. The nucleic acid molecules can be single stranded or double stranded. Strands can include the coding or non-coding strand.

The invention encompasses isolated or substantially purified nucleic acid or polypeptide compositions. An "isolated" or "purified" nucleic acid molecule or polypeptide, or biologically active portion thereof, is substantially free of other cellular material, or culture medium when produced by recombinant techniques, or substantially free of chemical precursors or other chemicals when chemically synthesized. Preferably, an "isolated" nucleic acid is free of sequences (preferably polypeptide encoding sequences) that naturally flank the nucleic acid (i.e., sequences located at the 5' and 3' ends of the nucleic acid) in the genomic DNA of the organism from which the nucleic acid is derived. For example, in various embodiments, the isolated nucleic acid molecule can contain less than about 5 kb, 4 kb, 3 kb, 2 kb, 1 kb, 0.5 kb, or 0.1 kb of nucleotide sequences that naturally flank the nucleic acid molecule in genomic DNA of the cell from which the nucleic acid is derived. A polypeptide that is substantially free of cellular material includes preparations of polypeptide having less than about 30%, 20%, 10%, 5%, (by dry weight) of contaminating polypeptide. When the polypeptide of the invention or biologically active portion thereof is recombinantly produced, preferably culture medium represents less than about 30%, 20%, 10%, or 5% (by dry weight) of chemical precursors or non-polypeptide-of-interest chemicals.

It is understood, however, that there are embodiments in which preparations that do not contain the substantially pure polypeptide may also be useful. Thus, less pure preparations can be useful where the contaminating material does not interfere with the specific desired use of the peptide. The compositions of the invention also encompass fragments and variants of the disclosed nucleotide sequences and the polypeptides encoded thereby.

The compositions of the invention are useful for, among other uses, expressing the receptor polypeptides in cells of interest to produce cellular or isolated preparations of the polypeptides for investigating the structure-function relationships of

*Bt* toxin receptors; investigating the toxin-receptor interactions; elucidating the mode of action of *Bt* toxins; screening and identifying novel *Bt* toxin receptor ligands including novel insecticidal toxins; and designing and developing novel *Bt* toxin receptor ligands including novel insecticidal toxins.

5           The isolated nucleotide sequences encoding the receptor polypeptides of the invention are expressed in a cell of interest; and the *Bt* toxin receptor polypeptides produced by the expression is utilized in intact cell or *in-vitro* receptor binding assays, and/or intact cell toxicity assays. Methods and conditions for *Bt* toxin binding and toxicity assays are known in the art and include but are not limited to those described  
10 in United States Patent NO: 5,693,491; T.P. Keeton *et al.* (1998) *Appl. Environ. Microbiol.* 64(6):2158-2165; B.R. Francis *et al.* (1997) *Insect Biochem. Mol. Biol.* 27(6):541-550; T.P. Keeton *et al.* (1997) *Appl. Environ. Microbiol.* 63(9):3419-3425; R.K. Vadlamudi *et al.* (1995) *J. Biol. Chem.* 270(10):5490-5494; Ihara *et al.* (1998) *Comparative Biochem. Physiol. B* 120:197-204; Nagamatsu *et al.* (1998) *Biosci.*  
15 *Biotechnol. Biochem.* 62(4):727-734, herein incorporated by reference. Such methods could be modified by one of ordinary skill in the art to develop assays utilizing the polypeptides of the invention.

          By "cell of interest" is intended any cell in which expression of the polypeptides of the invention is desired. Cells of interest include, but are not limited to mammalian,  
20 avian, insect, plant, bacteria, fungi and yeast cells. Cells of interest include but are not limited to cultured cell lines, primary cell cultures, cells *in vivo*, and cells of transgenic organisms.

          The methods of the invention encompass using the polypeptides encoded by the nucleotide sequences of the invention in receptor binding and/or toxicity assays to  
25 screen candidate ligands and identify novel *Bt* toxin receptor ligands, including receptor agonists and antagonists. Candidate ligands include molecules available from diverse libraries of small molecules created by combinatorial synthetic methods. Candidate ligands also include, but are not limited to antibodies, peptides, and other small molecules designed or deduced to interact with the receptor polypeptides of the  
30 invention. Candidate ligands include but are not limited to peptide fragments of the receptor, anti-receptor antibodies, anti-idiotypic antibodies mimicking one or more receptor binding domains of a toxin, fusion proteins produced by combining two or more toxins or fragments thereof, and the like. Ligands identified by the screening

methods of the invention include potential novel insecticidal toxins, the insecticidal activity of which can be determined by known methods; for example, as described in U.S. Patent No: 5,407,454; U.S. Application NO: 09/218,942; U.S. Application No: 09/003,217.

5       The invention provides methods for screening for ligands that bind to the polypeptides described herein. Both the polypeptides and relevant fragments thereof (for example, the toxin binding domain) can be used to screen by assay for compounds that bind to the receptor and exhibit desired binding characteristics. Desired binding characteristics include, but are not limited to binding affinity, binding site specificity, 10 association and dissociation rates, and the like. The screening assays could be intact cell or *in vitro* assays which include exposing a ligand binding domain to a sample ligand and detecting the formation of a ligand-binding polypeptide complex. The assays could be direct ligand-receptor binding assays or ligand competition assays.

      In one embodiment, the methods comprise providing at least one *Bt* toxin 15 receptor polypeptide of the invention, contacting the polypeptide with a sample and a control ligand under conditions promoting binding; and determining binding characteristics of sample ligands, relative to control ligands. The methods encompass any method known to the skilled artisan which can be used to provide the polypeptides of the invention in a binding assay. For *in vitro* binding assays, the 20 polypeptide may be provided as isolated, lysed, or homogenized cellular preparations. Isolated polypeptides may be provided in solution, or immobilized to a matrix. Methods for immobilizing polypeptides are well known in the art, and include but are not limited to construction and use of fusion polypeptides with commercially available high affinity ligands. For example, GST fusion proteins can be adsorbed 25 onto glutathione sepharose beads (Sigma Chemical, St. Louis, MO) or glutathione derivatized microtitre plates. The polypeptides can also be immobilized utilizing well techniques in the art utilizing conjugation of biotin and streptavidin. The polypeptides can also be immobilized utilizing well known techniques in the art utilizing chemical conjugation (linking) of polypeptides to a matrix. Alternatively, the 30 polypeptides may be provided in intact cell binding assays in which the polypeptides are generally expressed as cell surface *Bt* toxin receptors.

      The invention provides methods utilizing intact cell toxicity assays to screen for ligands that bind to the receptor polypeptides described herein and confer toxicity upon a

cell of interest expressing the polypeptide. A ligand selected by this screening is a potential insecticidal toxin to insects expressing the receptor polypeptides, particularly enterally. This deduction is premised on theories that insect specificity of a particular Bt toxin is determined by the presence of the receptor in specific insect species, or that binding of the toxins is specific for the receptor of some insect species and is insignificant or nonspecific for other variant receptors. See, for example Hofte *et al* (1989) *Microbiol Rev* 53: 242-255. The toxicity assays include exposing, in intact cells expressing a polypeptide of the invention, the toxin binding domain of the polypeptide to a sample ligand and detecting the toxicity effected in the cell expressing the polypeptide. By "toxicity" is intended the decreased viability of a cell. By "viability" is intended the ability of a cell to proliferate and/or differentiate and/or maintain its biological characteristics in a manner characteristic of that cell in the absence of a particular cytotoxic agent.

In one embodiment, the methods of the present invention comprise providing at least one cell surface Bt toxin receptor polypeptide of the invention comprising an extracellular toxin binding domain, contacting the polypeptide with a sample and a control ligand under conditions promoting binding, and determining the viability of the cell expressing the cell surface Bt toxin receptor polypeptide, relative to the control ligand.

By "contacting" is intended that the sample and control agents are presented to the intended ligand binding site of the polypeptides of the invention.

By "conditions promoting binding" is intended any combination of physical and biochemical conditions that enables a ligand of the polypeptides of the invention to determinably bind the intended polypeptide over background levels. Examples of such conditions for binding of Cry1 toxins to Bt toxin receptors, as well as methods for assessing the binding, are known in the art and include but are not limited to those described in Keeton *et al.* (1998) *Appl Environ Microbiol* 64(6): 2158-2165; Francis *et al.* (1997) *Insect Biochem Mol Biol* 27(6):541-550; Keeton *et al.* (1997) *Appl Environ Microbiol* 63(9):3419-3425; Vadlamudi *et al.* (1995) *J Biol Chem* 270(10):5490-5494; Ihara *et al.* (1998) *Comparative Biochemistry and Physiology, Part B* 120:197-204; and Nagamatsu *et al.* (1998) *Biosci. Biotechnol. Biochem.* 62(4):727-734, the contents of which are herein incorporated by reference. In this aspect of the present invention, known and commercially available methods for

studying protein-protein interactions, such as yeast and/or bacterial two-hybrid systems could also be used. Two-hybrid systems are available from, for example, CLONTECH (Palo Alto, Ca) or Display Systems Biotech Inc. (Vista, Ca).

The compositions and screening methods of the invention are useful for designing and developing novel *Bt* toxin receptor ligands including novel insecticidal toxins. Various candidate ligands; ligands screened and characterized for binding, toxicity, and species specificity; and/or ligands having known characteristics and specificities, could be linked or modified to produce novel ligands having particularly desired characteristics and specificities. The methods described herein for assessing binding, toxicity and insecticidal activity could be used to screen and characterize the novel ligands.

In one embodiment of the present invention, the sequences encoding the receptors of the invention, and variants and fragments thereof, are used with yeast and bacterial two-hybrid systems to screen for *Bt* toxins of interest (for example, more specific and/or more potent toxins), or for insect molecules that bind the receptor and can be used in developing novel insecticides.

By "linked" is intended that a covalent bond is produced between two or more molecules. Known methods that can be used for modification and/or linking of polypeptide ligands such as toxins, include but are not limited to mutagenic and recombinogenic approaches including but not limited to site-directed mutagenesis, chimeric polypeptide construction and DNA shuffling. Such methods are described in further detail below. Known polypeptide modification methods also include methods for covalent modification of polypeptides. "Operably linked" means that the linked molecules carry out the function intended by the linkage.

The compositions and screening methods of the present invention are useful for targeting ligands to cells expressing the receptor polypeptides of the invention. For targeting, secondary polypeptides, and/or small molecules which do not bind the receptor polypeptides of the invention are linked with one or more primary ligands which bind the receptor polypeptides; including but not limited to Cry1A toxin; more particularly Cry1 A(b) toxin or a fragment thereof. By this linkage, any polypeptide and/or small molecule linked to a primary ligand could be targeted to the receptor polypeptide, and thereby to a cell expressing the receptor polypeptide; wherein the ligand binding site is available at the extracellular surface of the cell.

In one embodiment of the invention, at least one secondary polypeptide toxin is linked with a primary Cry1 A toxin capable of binding the receptor polypeptides of the invention to produce a combination toxin which is targeted and toxic to insects expressing the receptor for the primary toxin. Such insects include those of the order  
5 *lepidoptera*, superfamily *Pyraloidea* and particularly from the species *Ostrinia*, specifically *Ostrinia nubilalis*. Such insects include the lepidopterans *S. frugiperda* and *H. Zea*. Such a combination toxin is particularly useful for eradicating or reducing crop damage by insects which have developed resistance to the primary toxin.

10 For expression of the *Bt* toxin receptor polypeptides of the invention in a cell of interest, the *Bt* toxin receptor sequences are provided in expression cassettes. The cassette will include 5' and 3' regulatory sequences operably linked to a *Bt* toxin receptor sequence of the invention. In this aspect of the present invention, by "operably linked" is intended a functional linkage between a promoter and a second  
15 sequence, wherein the promoter sequence initiates and mediates transcription of the DNA sequence corresponding to the second sequence. In reference to nucleic acids, generally, operably linked means that the nucleic acid sequences being linked are contiguous and, where necessary to join two polypeptide coding regions, contiguous and in the same reading frame. The cassette may additionally contain at least one  
20 additional gene to be cotransformed into the organism. Alternatively, the additional gene(s) can be provided on multiple expression cassettes.

Such an expression cassette is provided with a plurality of restriction sites for insertion of the *Bt* toxin receptor sequence to be under the transcriptional regulation of the regulatory regions. The expression cassette may additionally contain selectable  
25 marker genes.

The expression cassette will include in the 5'-3' direction of transcription, a transcriptional and translational initiation region, a *Bt* toxin receptor nucleotide sequence of the invention, and a transcriptional and translational termination region functional in host cells. The transcriptional initiation region, the promoter, may be  
30 native or analogous, or foreign or heterologous to the plant host. Additionally, the promoter may be the natural sequence or alternatively a synthetic sequence. By "foreign" is intended that the transcriptional initiation region is not found in the native host cells into which the transcriptional initiation region is introduced. As used

herein, a chimeric gene comprises a coding sequence operably linked to a transcription initiation region that is heterologous to the coding sequence.

While it may be preferable to express the sequences using heterologous promoters, the native promoter sequences may be used. Such constructs would  
5 change expression levels of *Bt* toxin receptor in the cell of interest. Thus, the phenotype of the cell is altered.

The termination region may be native with the transcriptional initiation region, may be native with the operably linked DNA sequence of interest, or may be derived from another source.

10 Where appropriate, the gene(s) may be optimized for increased expression in a particular transformed cell of interest. That is, the genes can be synthesized using host cell-preferred codons for improved expression.

Additional sequence modifications are known to enhance gene expression in a cellular host. These include elimination of sequences encoding spurious  
15 polyadenylation signals, exon-intron splice site signals, transposon-like repeats, and other such well-characterized sequences that may be deleterious to gene expression. The G-C content of the sequence may be adjusted to levels average for a given cellular host, as calculated by reference to known genes expressed in the host cell. When possible, the sequence is modified to avoid predicted hairpin secondary mRNA  
20 structures.

The expression cassettes may additionally contain 5' leader sequences in the expression cassette construct. Such leader sequences can act to enhance translation. Translation leaders are known in the art and include: picornavirus leaders, for example, EMCV leader (Encephalomyocarditis 5' noncoding region) (Elroy-Stein *et al.*  
25 *al.* (1989) *PNAS USA* 86:6126-6130); potyvirus leaders, for example, TEV leader (Tobacco Etch Virus) (Allison *et al.* (1986); MDMV leader (Maize Dwarf Mosaic Virus); *Virology* 154:9-20); and human immunoglobulin heavy-chain binding polypeptide (BiP), (Macejak *et al.* (1991) *Nature* 353:90-94); untranslated leader from the coat polypeptide mRNA of alfalfa mosaic virus (AMV RNA 4) (Jobling *et al.*  
30 (1987) *Nature* 325:622-625); tobacco mosaic virus leader (TMV) (Gallie *et al.* (1989) in *Molecular Biology of RNA*, ed. Cech (Liss, New York), pp. 237-256); and maize chlorotic mottle virus leader (MCMV) (Lommel *et al.* (1991) *Virology* 81:382-385).

See also, Della-Cioppa *et al.* (1987) *Plant Physiol.* 84:965-968. Other methods known to enhance translation can also be utilized, for example, introns, and the like.

In preparing the expression cassette, the various DNA fragments may be manipulated, so as to provide for the DNA sequences in the proper orientation and, as appropriate, in the proper reading frame. Toward this end, adapters or linkers may be employed to join the DNA fragments or other manipulations may be involved to provide for convenient restriction sites, removal of superfluous DNA, removal of restriction sites, or the like. For this purpose, *in vitro* mutagenesis, primer repair, restriction, annealing, resubstitutions, e.g., transitions and transversions, may be involved.

Using the nucleic acids of the present invention, the polypeptides of the invention could be expressed in any cell of interest, the particular choice of the cell depending on factors such as the level of expression and/or receptor activity desired. Cells of interest include, but are not limited to conveniently available mammalian, plant, insect, bacteria, and yeast host cells. The choice of promoter, terminator, and other expression vector components will also depend on the cell chosen. The cells produce the protein in a non-natural condition (e.g., in quantity, composition, location, and/or time), because they have been genetically altered through human intervention to do so.

It is expected that those of skill in the art are knowledgeable in the numerous expression systems available for expression of a nucleic acid encoding a protein of the present invention. No attempt to describe in detail the various methods known for the expression of proteins in prokaryotes or eukaryotes will be made.

In brief summary, the expression of isolated nucleic acids encoding a protein of the present invention will typically be achieved by operably linking, for example, the DNA or cDNA to a promoter, followed by incorporation into an expression vector. The vectors can be suitable for replication and integration in either prokaryotes or eukaryotes. Typical expression vectors contain transcription and translation terminators, initiation sequences, and promoters useful for regulation of the expression of the DNA encoding a protein of the present invention. To obtain high level expression of a cloned gene, it is desirable to construct expression vectors which contain, at the minimum, a strong promoter to direct transcription, a ribosome binding site for translational initiation, and a transcription/translation terminator. One



of skill would recognize that modifications can be made to a protein of the present invention without diminishing its biological activity. Some modifications may be made to facilitate the cloning, expression, or incorporation of the targeting molecule into a fusion protein. Such modifications are well known to those of skill in the art and include, for example, a methionine added at the amino terminus to provide an initiation site, or additional amino acids (e.g., poly His) placed on either terminus to create conveniently located restriction sites or termination codons or purification sequences.

Prokaryotic cells may be used as hosts for expression. Prokaryotes most frequently are represented by various strains of *E. coli*; however, other microbial strains may also be used. Commonly used prokaryotic control sequences which are defined herein to include promoters for transcription initiation, optionally with an operator, along with ribosome binding site sequences, include such commonly used promoters as the beta lactamase (penicillinase) and lactose (lac) promoter systems (Chang *et al.* (1977) *Nature* 198:1056), the tryptophan (trp) promoter system (Goeddel *et al.* (1980) *Nucleic Acids Res.* 8:4057) and the lambda-derived P<sub>L</sub> promoter and N-gene ribosome binding site (Shimatake *et al.* (1981) *Nature* 292:128). The inclusion of selection markers in DNA vectors transfected in *E. coli* is also useful. Examples of such markers include genes specifying resistance to ampicillin, tetracycline, or chloramphenicol.

The vector is selected to allow introduction into the appropriate host cell. Bacterial vectors are typically of plasmid or phage origin. Appropriate bacterial cells are infected with phage vector particles or transfected with naked phage vector DNA. If a plasmid vector is used, the bacterial cells are transfected with the plasmid vector DNA. Expression systems for expressing a protein of the present invention are available using *Bacillus sp.* and *Salmonella* (Palva *et al.* (1983) *Gene* 22:229-235; Mosbach *et al.* (1983) *Nature* 302:543-545).

A variety of eukaryotic expression systems such as yeast, insect cell lines, plant and mammalian cells, are known to those of skill in the art. The sequences of the present invention can be expressed in these eukaryotic systems. In some embodiments, transformed/transfected plant cells are employed as expression systems for production of the proteins of the instant invention.

Synthesis of heterologous proteins in yeast is well known. Sherman, F. *et al.* (1982) *Methods in Yeast Genetics*, Cold Spring Harbor Laboratory is a well recognized work describing the various methods available to produce the protein in yeast. Two widely utilized yeast for production of eukaryotic proteins are

5 *Saccharomyces cerevisia* and *Pichia pastoris*. Vectors, strains, and protocols for expression in *Saccharomyces* and *Pichia* are known in the art and available from commercial suppliers (e.g., Invitrogen). Suitable vectors usually have expression control sequences, such as promoters, including 3-phosphoglycerate kinase or alcohol oxidase, and an origin of replication, termination sequences and the like as desired.

10 A protein of the present invention, once expressed, can be isolated from yeast by lysing the cells and applying standard protein isolation techniques to the lysates. The monitoring of the purification process can be accomplished by using Western blot techniques or radioimmunoassay or other standard immunoassay techniques.

The sequences encoding proteins of the present invention can also be ligated

15 to various expression vectors for use in transfecting cell cultures of, for instance, mammalian, insect, or plant origin. Illustrative of cell cultures useful for the production of the peptides are mammalian cells. Mammalian cell systems often will be in the form of monolayers of cells although mammalian cell suspensions may also be used. A number of suitable host cell lines capable of expressing intact proteins

20 have been developed in the art, and include the COS, HEK293, BHK21, and CHO cell lines. Expression vectors for these cells can include expression control sequences, such as an origin of replication, a promoter (e.g., the CMV promoter, a HSV *tk* promoter or *pgk* (phosphoglycerate kinase promoter)), an enhancer (Queen *et al.* (1986) *Immunol. Rev.* 89:49), and necessary processing information sites, such as

25 ribosome binding sites, RNA splice sites, polyadenylation sites (e.g., an SV40 large T Ag poly A addition site), and transcriptional terminator sequences. Other animal cells useful for production of proteins of the present invention are available, for instance, from the American Type Culture Collection Catalogue of Cell Lines and Hybridomas (7th edition, 1992). A particular example of mammalian cells for expression of a Bt

30 toxin receptor and assessing Bt toxin cytotoxicity mediated by the receptor, includes embryonic 293 cells. See U.S. Patent NO. 5,693,491, herein incorporated by reference.

Appropriate vectors for expressing proteins of the present invention in insect cells are usually derived from the SF9 baculovirus. Suitable insect cell lines include mosquito larvae, silkworm, armyworm, moth and *Drosophila* cell lines such as a Schneider cell line (See Schneider *et al.* (1987) *J. Embryol. Exp. Morphol.* 27: 353-365).

As with yeast, when higher animal or plant host cells are employed, polyadenylation or transcription terminator sequences are typically incorporated into the vector. An example of a terminator sequence is the polyadenylation sequence from the bovine growth hormone gene. Sequences for accurate splicing of the transcript may also be included. An example of a splicing sequence is the VP1 intron from SV40 (Sprague *et al.* (1983) *J. Virol.* 45:773-781). Additionally, gene sequences to control replication in the host cell may be incorporated into the vector such as those found in bovine papilloma virus-type vectors. Saveria-Campo, M., Bovine Papilloma Virus DNA a Eukaryotic Cloning Vector in *DNA Cloning Vol. II a Practical Approach*, D.M. Glover, ed., IRL Pres, Arlington, Virginia pp. 213-238 (1985).

In a particular embodiment of the invention, it may be desirable to negatively control receptor binding; particularly, when toxicity to a cell is no longer desired or if it is desired to reduce toxicity to a lower level. In this case, ligand-receptor polypeptide binding assays can be used to screen for compounds which bind to the receptor but do not confer toxicity to a cell expressing the receptor. The examples of a molecule that can be used to block ligand binding include an antibody that specifically recognizes the ligand binding domain of the receptor such that ligand binding is decreased or prevented as desired.

In another embodiment, receptor polypeptide expression could be blocked by the use of antisense molecules directed against receptor RNA or ribozymes specifically targeted to this receptor RNA. It is recognized that with the provided nucleotide sequences, antisense constructions, complementary to at least a portion of the messenger RNA (mRNA) for the *Bt* toxin receptor sequences can be constructed. Antisense nucleotides are constructed to hybridize with the corresponding mRNA. Modifications of the antisense sequences may be made as long as the sequences hybridize to and interfere with expression of the corresponding mRNA. In this manner, antisense constructions having 70%, preferably 80%, more preferably 85%

sequence similarity to the corresponding antisensed sequences may be used. Furthermore, portions of the antisense nucleotides may be used to disrupt the expression of the target gene. Generally, sequences of at least 50 nucleotides, 100 nucleotides, 200 nucleotides, or greater may be used.

5           Fragments and variants of the disclosed nucleotide sequences and polypeptides encoded thereby are encompassed by the present invention. By "fragment" is intended a portion of the nucleotide sequence, or a portion of the amino acid sequence, and hence a portion of the polypeptide encoded thereby. Fragments of a nucleotide sequence may encode polypeptide fragments that retain the biological  
10   activity of the native polypeptide and, for example, bind *Bt* toxins. Alternatively, fragments of a nucleotide sequence that are useful as hybridization probes generally do not encode fragment polypeptides retaining biological activity. Thus, fragments of a nucleotide sequence may range from at least about 20 nucleotides, about 50  
15   nucleotides, about 100 nucleotides, and up to the full-length nucleotide sequence encoding the polypeptides of the invention.

          A fragment of a *Bt* toxin receptor nucleotide sequence that encodes a biologically active portion of a *Bt* toxin receptor polypeptide of the invention will encode at least 15, 25, 30, 50, 100, 150, 200 or 250 contiguous amino acids, or up to the total number of amino acids present in a full-length *Bt* toxin receptor polypeptide  
20   of the invention (for example, 1717, 1730, and 1734 amino acids for SEQ ID NOs:2, 4, and 6, respectively. Fragments of a *Bt* toxin receptor nucleotide sequence that are useful as hybridization probes for PCR primers generally need not encode a biologically active portion of a *Bt* toxin receptor polypeptide.

          Thus, a fragment of a *Bt* toxin receptor nucleotide sequence may encode a  
25   biologically active portion of a *Bt* toxin receptor polypeptide, or it may be a fragment that can be used as a hybridization probe or PCR primer using methods disclosed below. A biologically active portion of a *Bt* toxin receptor polypeptide can be prepared by isolating a portion of one of the *Bt* toxin receptor nucleotide sequences of the invention, expressing the encoded portion of the *Bt* toxin receptor polypeptide  
30   (e.g., by recombinant expression *in vitro*), and assessing the activity of the encoded portion of the *Bt* toxin receptor polypeptide. Nucleic acid molecules that are fragments of a *Bt* toxin receptor nucleotide sequence comprise at least 16, 20, 50, 75, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 800, 900, 1,000,

1,100, 1,200, 1,300, or 1,400 nucleotides, or up to the number of nucleotides present in a full-length *Bt* toxin receptor nucleotide sequence disclosed herein (for example, 5498, 5527, and 5614 nucleotides for SEQ ID NOs: 1, 3, and 5, respectively).

By "variants" is intended substantially similar sequences. For nucleotide  
5 sequences, conservative variants include those sequences that, because of the degeneracy of the genetic code, encode the amino acid sequence of one of the *Bt* toxin receptor polypeptides of the invention. Naturally occurring allelic variants such as these can be identified with the use of well-known molecular biology techniques, as, for example, with polymerase chain reaction (PCR) and hybridization techniques as  
10 outlined below. Variant nucleotide sequences also include synthetically derived nucleotide sequences, such as those generated, for example, by using site-directed mutagenesis, but which still encode a *Bt* toxin receptor protein of the invention. Generally, variants of a particular nucleotide sequence of the invention will have at least about 40%, 50%, 60%, 65%, 70%, generally at least about 75%, 80%, 85%,  
15 preferably at least about 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, and more preferably at least about 98%, 99% or more sequence identity to that particular nucleotide sequence as determined by sequence alignment programs described elsewhere herein using default parameters.

By "variant" protein is intended a protein derived from the native protein by  
20 deletion (so-called truncation) or addition of one or more amino acids to the N-terminal and/or C-terminal end of the native protein; deletion or addition of one or more amino acids at one or more sites in the native protein; or substitution of one or more amino acids at one or more sites in the native protein. Variant proteins encompassed by the present invention are biologically active, that is they continue to  
25 possess the desired biological activity of the native protein, that is, activity as described herein (for example, *Bt* toxin binding activity). Such variants may result from, for example, genetic polymorphism or from human manipulation. Biologically active variants of a native *Bt* toxin receptor protein of the invention will have at least about 40%, 50%, 60%, 65%, 70%, generally at least about 75%, 80%, 85%,  
30 preferably at least about 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, and more preferably at least about 98%, 99% or more sequence identity to the amino acid sequence for the native protein as determined by sequence alignment programs described elsewhere herein using default parameters. A biologically active variant of a

protein of the invention may differ from that protein by as few as 1-15 amino acid residues, as few as 1-10, such as 6-10, as few as 5, as few as 4, 3, 2, or even 1 amino acid residue.

The polypeptides of the invention may be altered in various ways including amino acid substitutions, deletions, truncations, and insertions. Methods for such manipulations are generally known in the art. For example, amino acid sequence variants of the *Bt* toxin receptor polypeptides can be prepared by mutations in the DNA. Methods for mutagenesis and nucleotide sequence alterations are well known in the art. See, for example, Kunkel (1985) *Proc. Natl. Acad. Sci. USA* 82:488-492; Kunkel *et al.* (1987) *Methods in Enzymol.* 154:367-382; US Patent No. 4,873,192; Walker and Gastra, eds. (1983) *Techniques in Molecular Biology* (MacMillan Publishing Company, New York) and the references cited therein. Guidance as to appropriate amino acid substitutions that do not affect biological activity of the protein of interest may be found in the model of Dayhoff *et al.* (1978) *Atlas of Protein Sequence and Structure* (Natl. Biomed. Res. Found., Washington, D.C.), herein incorporated by reference. Conservative substitutions, such as exchanging one amino acid with another having similar properties, may be preferable.

Thus, the genes and nucleotide sequences of the invention include both the naturally occurring sequences as well as mutant forms. Likewise, the proteins of the invention encompass both naturally occurring proteins as well as variations and modified forms thereof. Such variants will continue to possess the desired toxin binding activity. Obviously, the mutations that will be made in the DNA encoding the variant must not place the sequence out of reading frame and preferably will not create complementary regions that could produce secondary mRNA structure. See, EP Patent Application Publication No. 75,444.

The deletions, insertions, and substitutions of the protein sequences encompassed herein are not expected to produce radical changes in the characteristics of the protein.

For example, it is recognized that at least about 10, 20, 50, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, and up to 960 amino acids may be deleted from the N-terminus of a polypeptide that has the amino acid sequence set forth in SEQ ID NO:2, and still retain binding function. It is further recognized that at least about 10, 20, 30, 40, 50, 60, 70, 80, 90, 100, 110, and up to

119 amino acids may be deleted from the C-terminus of a polypeptide that has the amino acid sequence set forth in SEQ ID NO:2, and still retain binding function. Deletion variants of the invention that encompass polypeptides having these deletions. It is recognized that deletion variants of the invention that retain binding function  
5 encompass polypeptides having these N-terminal or C-terminal deletions, or having any deletion combination thereof at both the C- and the N-termini.

However, when it is difficult to predict the exact effect of the substitution, deletion, or insertion in advance of doing so, one skilled in the art will appreciate that the effect will be evaluated by routine screening assays. That is, the activity can be  
10 evaluated by receptor binding and/or toxicity assays. See, for example, United States Patent NO: 5,693,491; T.P. Keeton *et al.* (1998) *Appl. Environ. Microbiol.* 64(6):2158-2165; B.R. Francis *et al.* (1997) *Insect Biochem. Mol. Biol.* 27(6):541-550; T.P. Keeton *et al.* (1997) *Appl. Environ. Microbiol.* 63(9):3419-3425; R.K. Vadlamudi *et al.* (1995) *J. Biol. Chem.* 270(10):5490-5494; Ihara *et al.* (1998)  
15 *Comparative Biochem. Physiol. B* 120:197-204; Nagamatsu *et al.* (1998) *Biosci. Biotechnol. Biochem.* 62(4):727-734, herein incorporated by reference.

Variant nucleotide sequences and polypeptides also encompass sequences and polypeptides derived from a mutagenic and recombinogenic procedure such as DNA shuffling. With such a procedure, one or more different toxin receptor coding  
20 sequences can be manipulated to create a new toxin receptor, including but not limited to a new *Bt* toxin receptor; possessing the desired properties. In this manner, libraries of recombinant polynucleotides are generated from a population of related sequence polynucleotides comprising sequence regions that have substantial sequence identity and can be homologously recombined *in vitro* or *in vivo*. For example, using this  
25 approach, sequence motifs encoding a domain of interest may be shuffled between the *Bt* toxin receptor gene of the invention and other known *Bt* toxin receptor genes to obtain a new gene coding for a polypeptide with an improved property of interest, such as an increased ligand affinity in the case of a receptor. Strategies for such DNA shuffling are known in the art. See, for example, Stemmer (1994) *Proc. Natl. Acad.*  
30 *Sci. USA* 91:10747-10751; Stemmer (1994) *Nature* 370:389-391; Cramer *et al.* (1997) *Nature Biotech.* 15:436-438; Moore *et al.* (1997) *J. Mol. Biol.* 272:336-347; Zhang *et al.* (1997) *Proc. Natl. Acad. Sci. USA* 94:4504-4509; Cramer *et al.* (1998) *Nature* 391:288-291; and U.S. Patent Nos. 5,605,793 and 5,837,448.

Where the receptor polypeptides of the invention are expressed in a cell and associated with the cell membrane (for example, by a transmembrane segment), in order for the receptor of the invention to bind a desired ligand, for example a Cry I A toxin, the receptor's ligand binding domain must be available to the ligand. In this aspect, it is  
5 recognized that the native *Bt* toxin receptor of the invention is oriented such that the toxin binding site is available extracellularly.

Accordingly, in methods comprising use of intact cells, the invention provides cell surface *Bt*-toxin receptors. By a "cell surface *Bt* toxin receptor" is intended a membrane-bound receptor polypeptide comprising at least one extracellular *Bt* toxin  
10 binding site. A cell surface receptor of the invention comprises an appropriate combination of signal sequences and transmembrane segments for guiding and retaining the receptor at the cell membrane such that that toxin binding site is available extracellularly. Where native *Bt* toxin receptors are used for expression, deduction of the composition and configuration of the signal sequences and transmembrane segments is  
15 not necessary to ensure the appropriate topology of the polypeptide for displaying the toxin binding site extracellularly. As an alternative to native signal and transmembrane sequences, heterologous signal and transmembrane sequences could be utilized to produce a cell surface receptor polypeptide of the invention.

It is recognized that it may be of interest to generate *Bt* toxin receptors that are  
20 capable of interacting with the receptor's ligands intracellularly in the cytoplasm, in the nucleus or other organelles, in other subcellular spaces; or in the extracellular space. Accordingly, the invention encompasses variants of the receptors of the invention, wherein one or more of the segments of the receptor polypeptide is modified to target the polypeptide to a desired intra- or extracellular location.

Also encompassed by the invention are receptor fragments and variants that are  
25 useful, among other things, as binding antagonists that will compete with a cell surface receptor of the invention. Such a fragment or variant can, for example, bind a toxin but not be able to confer toxicity to a particular cell. In this aspect, the invention provides secreted receptors, more particularly secreted *Bt* toxin receptors; or receptors that are not  
30 membrane bound. The secreted receptors of the invention can contain a heterologous or homologous signal sequence facilitating its secretion from the cell expressing the receptors; and further comprise a secretion variation in the region corresponding to transmembrane segments. By "secretion variation" is intended that amino acids



corresponding to a transmembrane segment of a membrane bound receptor comprise one or more deletions, substitutions, insertions, or any combination thereof; such that the region no longer retains the requisite hydrophobicity to serve as a transmembrane segment. Sequence alterations to create a secretion variation can be tested by  
5 confirming secretion of the polypeptide comprising the variation from the cell expressing the polypeptide.

The polypeptides of the invention can be purified from cells that naturally express it, purified from cells that have been altered to express it (i. e. recombinant) or synthesized using polypeptide synthesis techniques that are well known in the art. In one  
10 embodiment, the polypeptide is produced by recombinant DNA methods. In such methods a nucleic acid molecule encoding the polypeptide is cloned into an expression vector as described more fully herein and expressed in an appropriate host cell according to known methods in the art. The polypeptide is then isolated from cells using polypeptide purification techniques well known to those of ordinary skill in the art.  
15 Alternatively, the polypeptide or fragment can be synthesized using peptide synthesis methods well known to those of ordinary skill in the art.

The invention also encompasses fusion polypeptides in which one or more polypeptides of the invention are fused with at least one polypeptide of interest. In one embodiment, the invention encompasses fusion polypeptides in which a heterologous  
20 polypeptide of interest has an amino acid sequence that is not substantially homologous to the polypeptide of the invention. In this embodiment, the polypeptide of the invention and the polypeptide of interest may or may not be operatively linked. An example of operative linkage is fusion in-frame so that a single polypeptide is produced upon translation. Such fusion polypeptides can, for example, facilitate the purification of a  
25 recombinant polypeptide.

In another embodiment, the fused polypeptide of interest may contain a heterologous signal sequence at the N-terminus facilitating its secretion from specific host cells. The expression and secretion of the polypeptide can thereby be increased by use of the heterologous signal sequence.

30 The invention is also directed to polypeptides in which one or more domains in the polypeptide described herein are operatively linked to heterologous domains having homologous functions. Thus, the toxin binding domain can be replaced with a toxin binding domain for other toxins. Thereby, the toxin specificity of the receptor is based

on a toxin binding domain other than the domain encoded by *Bt* toxin receptor but other characteristics of the polypeptide, for example, membrane localization and topology is based on *Bt* toxin receptor.

Alternatively, the native *Bt* toxin binding domain may be retained while  
5 additional heterologous ligand binding domains, including but not limited to heterologous toxin binding domains are comprised by the receptor. Thus, the invention also encompasses fusion polypeptides in which a polypeptide of interest is a heterologous polypeptide comprising a heterologous toxin binding domains. Examples of heterologous polypeptides comprising Cry1 toxin binding domains include, but are  
10 not limited to Knight et al (1994) *Mol Micro* 11: 429-436; Lee et al. (1996) *Appl Environ Micro* 63: 2845-2849; Gill et al. (1995) *J Biol Chem* 270: 27277-27282; Garczynski et al. (1991) *Appl Environ Microbiol* 10: 2816-2820; Vadlamudi et al. (1995) *J Biol Chem* 270(10):5490-4, U.S. Patent No5,693,491.

The *Bt* toxin receptor peptide of the invention may also be fused with other  
15 members of the cadherin superfamily. Such fusion polypeptides could provide an important reflection of the binding properties of the members of the superfamily. Such combinations could be further used to extend the range of applicability of these molecules in a wide range of systems or species that might not otherwise be amenable to native or relatively homologous polypeptides. The fusion constructs could be substituted  
20 into systems in which a native construct would not be functional because of species specific constraints. Hybrid constructs may further exhibit desirable or unusual characteristics otherwise unavailable with the combinations of native polypeptides.

Polypeptide variants encompassed by the present invention include those that contain mutations that either enhance or decrease one or more domain functions. For  
25 example, in the toxin binding domain, a mutation may be introduced that increases or decreases the sensitivity of the domain to a specific toxin.

As an alternative to the introduction of mutations, increase in function may be provided by increasing the copy number of ligand binding domains. Thus, the invention also encompasses receptor polypeptides in which the toxin binding domain is provided  
30 in more than one copy.

The invention further encompasses cells containing receptor expression vectors comprising the *Bt* toxin receptor sequences, and fragments and variants thereof. The expression vector can contain one or more expression cassettes used to transform a cell

of interest. Transcription of these genes can be placed under the control of a constitutive or inducible promoter (for example, tissue - or cell cycle-preferred).

Where more than one expression cassette utilized, the cassette that is additional to the cassette comprising at least one receptor sequence of the invention, can comprise  
5 either a receptor sequence of the invention or any other desired sequences.

The nucleotide sequences of the invention can be used to isolate homologous sequences in insect species other than *ostrinia*, particularly other lepidopteran species, more particularly other *Pyraloidea* species.

The following terms are used to describe the sequence relationships between  
10 two or more nucleic acids or polynucleotides: (a) "reference sequence", (b) "comparison window", (c) "sequence identity", (d) "percentage of sequence identity", and (e) "substantial identity".

(a) As used herein, "reference sequence" is a defined sequence used as a basis for sequence comparison. A reference sequence may be a subset or the entirety  
15 of a specified sequence; for example, as a segment of a full-length cDNA or gene sequence, or the complete cDNA or gene sequence.

(b) As used herein, "comparison window" makes reference to a contiguous and specified segment of a polynucleotide sequence, wherein the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps)  
20 compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. Generally, the comparison window is at least 20 contiguous nucleotides in length, and optionally can be 30, 40, 50, 100, or longer. Those of skill in the art understand that to avoid a high similarity to a reference sequence due to inclusion of gaps in the polynucleotide sequence a gap  
25 penalty is typically introduced and is subtracted from the number of matches.

Methods of alignment of sequences for comparison are well known in the art. Thus, the determination of percent identity between any two sequences can be accomplished using a mathematical algorithm. Non-limiting examples of such mathematical algorithms are the algorithm of Myers and Miller (1988) *CABIOS* 4:11-  
30 17; the local homology algorithm of Smith *et al.* (1981) *Adv. Appl. Math.* 2:482; the homology alignment algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48:443-453; the search-for-similarity-method of Pearson and Lipman (1988) *Proc. Natl. Acad. Sci.* 85:2444-2448; the algorithm of Karlin and Altschul (1990) *Proc.*

*Natl. Acad. Sci. USA* 87:2264, modified as in Karlin and Altschul (1993) *Proc. Natl. Acad. Sci. USA* 90:5873-5877.

Computer implementations of these mathematical algorithms can be utilized for comparison of sequences to determine sequence identity. Such implementations include, but are not limited to: CLUSTAL in the PC/Gene program (available from  
5 Intelligenetics, Mountain View, California); the ALIGN program (Version 2.0); the ALIGN PLUS program (version 3.0, copyright 1997); and GAP, BESTFIT, BLAST, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Version 8 (available from Genetics Computer Group (GCG), 575 Science Drive, Madison,  
10 Wisconsin, USA). Alignments using these programs can be performed using the default parameters. The CLUSTAL program is well described by Higgins *et al.* (1988) *Gene* 73:237-244 (1988); Higgins *et al.* (1989) *CABIOS* 5:151-153; Corpet *et al.* (1988) *Nucleic Acids Res.* 16:10881-90; Huang *et al.* (1992) *CABIOS* 8:155-65; and Pearson *et al.* (1994) *Meth. Mol. Biol.* 24:307-331. The ALIGN and the ALIGN  
15 PLUS programs are based on the algorithm of Myers and Miller (1988) *supra*. A PAM120 weight residue table, a gap length penalty of 12, and a gap penalty of 4 can be used with the ALIGN program when comparing amino acid sequences. The BLAST programs of Altschul *et al.* (1990) *J. Mol. Biol.* 215:403 are based on the algorithm of Karlin and Altschul (1990) *supra*. BLAST nucleotide searches can be  
20 performed with the BLASTN program, score = 100, wordlength = 12, to obtain nucleotide sequences homologous to a nucleotide sequence encoding a protein of the invention. BLAST protein searches can be performed with the BLASTX program, score = 50, wordlength = 3, to obtain amino acid sequences homologous to a protein or polypeptide of the invention. To obtain gapped alignments for comparison  
25 purposes, Gapped BLAST (in BLAST 2.0) can be utilized as described in Altschul *et al.* (1997) *Nucleic Acids Res.* 25:3389. Alternatively, PSI-BLAST (in BLAST 2.0) can be used to perform an iterated search that detects distant relationships between molecules. See Altschul *et al.* (1997) *supra*. When utilizing BLAST, Gapped BLAST, PSI-BLAST, the default parameters of the respective programs (e.g., BLASTN for  
30 nucleotide sequences, BLASTX for proteins) can be used. See <http://www.ncbi.nlm.nih.gov>. Alignment may also be performed manually by inspection.

Unless otherwise stated, sequence identity/similarity values provided herein refer to the value obtained using GAP Version 10 using the following parameters: % identity using GAP Weight of 50 and Length Weight of 3; % similarity using Gap Weight of 12 and Length Weight of 4, or any equivalent program. By "equivalent program" is intended any sequence comparison program that, for any two sequences in question, generates an alignment having identical nucleotide or amino acid residue matches and an identical percent sequence identity when compared to the corresponding alignment generated by the preferred program.

GAP uses the algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48: 443-453, to find the alignment of two complete sequences that maximizes the number of matches and minimizes the number of gaps. GAP considers all possible alignments and gap positions and creates the alignment with the largest number of matched bases and the fewest gaps. It allows for the provision of a gap creation penalty and a gap extension penalty in units of matched bases. GAP must make a profit of gap creation penalty number of matches for each gap it inserts. If a gap extension penalty greater than zero is chosen, GAP must, in addition, make a profit for each gap inserted of the length of the gap times the gap extension penalty. Default gap creation penalty values and gap extension penalty values in Version 10 of the Wisconsin Genetics Software Package for protein sequences are 8 and 2, respectively. For nucleotide sequences the default gap creation penalty is 50 while the default gap extension penalty is 3. The gap creation and gap extension penalties can be expressed as an integer selected from the group of integers consisting of from 0 to 200. Thus, for example, the gap creation and gap extension penalties can be 0, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65 or greater.

GAP presents one member of the family of best alignments. There may be many members of this family, but no other member has a better quality. GAP displays four figures of merit for alignments: Quality, Ratio, Identity, and Similarity. The Quality is the metric maximized in order to align the sequences. Ratio is the quality divided by the number of bases in the shorter segment. Percent Identity is the percent of the symbols that actually match. Percent Similarity is the percent of the symbols that are similar. Symbols that are across from gaps are ignored. A similarity is scored when the scoring matrix value for a pair of symbols is greater than or equal to 0.50, the similarity threshold. The scoring matrix used in Version 10 of the

Wisconsin Genetics Software Package is BLOSUM62 (see Henikoff and Henikoff (1989). *Proc. Natl. Acad. Sci. USA* 89:10915).

(c) As used herein, "sequence identity" or "identity" in the context of two nucleic acid or polypeptide sequences makes reference to the residues in the two sequences that are the same when aligned for maximum correspondence over a specified comparison window. When percentage of sequence identity is used in reference to proteins it is recognized that residue positions which are not identical often differ by conservative amino acid substitutions, where amino acid residues are substituted for other amino acid residues with similar chemical properties (e.g., charge or hydrophobicity) and therefore do not change the functional properties of the molecule. When sequences differ in conservative substitutions, the percent sequence identity may be adjusted upwards to correct for the conservative nature of the substitution. Sequences that differ by such conservative substitutions are said to have "sequence similarity" or "similarity". Means for making this adjustment are well known to those of skill in the art. Typically this involves scoring a conservative substitution as a partial rather than a full mismatch, thereby increasing the percentage sequence identity. Thus, for example, where an identical amino acid is given a score of 1 and a non-conservative substitution is given a score of zero, a conservative substitution is given a score between zero and 1. The scoring of conservative substitutions is calculated, e.g., as implemented in the program PC/GENE (Intelligenetics, Mountain View, California).

(d) As used herein, "percentage of sequence identity" means the value determined by comparing two optimally aligned sequences over a comparison window, wherein the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) as compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical nucleic acid base or amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison, and multiplying the result by 100 to yield the percentage of sequence identity.

(e)(i) The term "substantial identity" of polynucleotide sequences means that a polynucleotide comprises a sequence that has at least 70% sequence identity,

preferably at least 80%, more preferably at least 90%, and most preferably at least 95%, compared to a reference sequence using one of the alignment programs described using standard parameters. One of skill in the art will recognize that these values can be appropriately adjusted to determine corresponding identity of proteins encoded by two nucleotide sequences by taking into account codon degeneracy, amino acid similarity, reading frame positioning, and the like. Substantial identity of amino acid sequences for these purposes normally means sequence identity of at least 60%, more preferably at least 70%, 80%, 90%, and most preferably at least 95%.

Another indication that nucleotide sequences are substantially identical is if two molecules hybridize to each other under stringent conditions. Generally, stringent conditions are selected to be about 5°C lower than the thermal melting point ( $T_m$ ) for the specific sequence at a defined ionic strength and pH. However, stringent conditions encompass temperatures in the range of about 1°C to about 20°C lower than the  $T_m$ , depending upon the desired degree of stringency as otherwise qualified herein. Nucleic acids that do not hybridize to each other under stringent conditions are still substantially identical if the polypeptides they encode are substantially identical. This may occur, e.g., when a copy of a nucleic acid is created using the maximum codon degeneracy permitted by the genetic code. One indication that two nucleic acid sequences are substantially identical is when the polypeptide encoded by the first nucleic acid sequence is immunologically cross reactive with the polypeptide encoded by the second nucleic acid sequence.

(e)(ii) The term "substantial identity" in the context of a peptide indicates that a peptide comprises a sequence with at least 70% sequence identity to a reference sequence, preferably 80%, more preferably 85%, most preferably at least 90% or 95% sequence identity to the reference sequence over a specified comparison window. Preferably, optimal alignment is conducted using the homology alignment algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48:443-453. An indication that two peptide sequences are substantially identical is that one peptide is immunologically reactive with antibodies raised against the second peptide. Thus, a peptide is substantially identical to a second peptide, for example, where the two peptides differ only by a conservative substitution. Peptides that are "substantially similar" share sequences as noted above except that residue positions that are not identical may differ by conservative amino acid changes.

The nucleotide sequences of the invention can be used to isolate corresponding sequences from other organisms, particularly other insects, more particularly other lepidopteran species. In this manner, methods such as PCR, hybridization, and the like can be used to identify such sequences based on their sequence homology to the sequences set forth herein. Sequences isolated based on their sequence identity to the entire *Bt* toxin receptor sequences set forth herein or to fragments thereof are encompassed by the present invention. Such sequences include sequences that are orthologs of the disclosed sequences. By "orthologs" is intended genes derived from a common ancestral gene and which are found in different species as a result of speciation. Genes found in different species are considered orthologs when their nucleotide sequences and/or their encoded protein sequences share substantial identity as defined elsewhere herein. Functions of orthologs are often highly conserved among species.

In a PCR approach, oligonucleotide primers can be designed for use in PCR reactions to amplify corresponding DNA sequences from cDNA or genomic DNA extracted from any organism of interest. Methods for designing PCR primers and PCR cloning are generally known in the art and are disclosed in Sambrook *et al.* (1989) *Molecular Cloning: A Laboratory Manual* (2d ed., Cold Spring Harbor Laboratory Press, Plainview, New York). See also Innis *et al.*, eds. (1990) *PCR Protocols: A Guide to Methods and Applications* (Academic Press, New York); Innis and Gelfand, eds. (1995) *PCR Strategies* (Academic Press, New York); and Innis and Gelfand, eds. (1999) *PCR Methods Manual* (Academic Press, New York). Known methods of PCR include, but are not limited to, methods using paired primers, nested primers, single specific primers, degenerate primers, gene-specific primers, vector-specific primers, partially-mismatched primers, and the like.

In hybridization techniques, all or part of a known nucleotide sequence is used as a probe that selectively hybridizes to other corresponding nucleotide sequences present in a population of cloned genomic DNA fragments or cDNA fragments (i.e., genomic or cDNA libraries) from a chosen organism. The hybridization probes may be genomic DNA fragments, cDNA fragments, RNA fragments, or other oligonucleotides, and may be labeled with a detectable group such as  $^{32}\text{P}$ , or any other detectable marker. Thus, for example, probes for hybridization can be made by labeling synthetic oligonucleotides based on the *Bt* toxin receptor sequences of the



invention. Methods for preparation of probes for hybridization and for construction of cDNA and genomic libraries are generally known in the art and are disclosed in Sambrook *et al.* (1989) *Molecular Cloning: A Laboratory Manual* (2d ed., Cold Spring Harbor Laboratory Press, Plainview, New York).

5 For example, the entire *Bt* toxin receptor sequence disclosed herein, or one or more portions thereof, may be used as a probe capable of specifically hybridizing to corresponding *Bt* toxin receptor sequences and messenger RNAs. To achieve specific hybridization under a variety of conditions, such probes include sequences that are unique among *Bt* toxin receptor sequences and are preferably at least about 10  
10 nucleotides in length, and most preferably at least about 20 nucleotides in length. Such probes may be used to amplify corresponding *Bt* toxin receptor sequences from a chosen plant organism by PCR. This technique may be used to isolate additional coding sequences from a desired organism or as a diagnostic assay to determine the presence of coding sequences in an organism. Hybridization techniques include  
15 hybridization screening of plated DNA libraries (either plaques or colonies; see, for example, Sambrook *et al.* (1989) *Molecular Cloning: A Laboratory Manual* (2d ed., Cold Spring Harbor Laboratory Press, Plainview, New York).

Hybridization of such sequences may be carried out under stringent conditions. By "stringent conditions" or "stringent hybridization conditions" is  
20 intended conditions under which a probe will hybridize to its target sequence to a detectably greater degree than to other sequences (e.g., at least 2-fold over background). Stringent conditions are sequence-dependent and will be different in different circumstances. By controlling the stringency of the hybridization and/or washing conditions, target sequences that are 100% complementary to the probe can  
25 be identified (homologous probing). Alternatively, stringency conditions can be adjusted to allow some mismatching in sequences so that lower degrees of similarity are detected (heterologous probing). Generally, a probe is less than about 1000 nucleotides in length, preferably less than 500 nucleotides in length.

Typically, stringent conditions will be those in which the salt concentration is  
30 less than about 1.5 M Na ion, typically about 0.01 to 1.0 M Na ion concentration (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30°C for short probes (e.g., 10 to 50 nucleotides) and at least about 60°C for long probes (e.g., greater than 50 nucleotides). Stringent conditions may also be achieved with the addition of

destabilizing agents such as formamide. Exemplary low stringency conditions include hybridization with a buffer solution of 30 to 35% formamide, 1 M NaCl, 1% SDS (sodium dodecyl sulphate) at 37°C, and a wash in 1X to 2X SSC (20X SSC = 3.0 M NaCl/0.3 M trisodium citrate) at 50 to 55°C. Exemplary moderate stringency conditions include hybridization in 40 to 45% formamide, 1.0 M NaCl, 1% SDS at 37°C, and a wash in 0.5X to 1X SSC at 55 to 60°C. Exemplary high stringency conditions include hybridization in 50% formamide, 1 M NaCl, 1% SDS at 37°C, and a wash in 0.1X SSC at 60 to 65°C. Duration of hybridization is generally less than about 24 hours, usually about 4 to about 12 hours.

Specificity is typically the function of post-hybridization washes, the critical factors being the ionic strength and temperature of the final wash solution. For DNA-DNA hybrids, the  $T_m$  can be approximated from the equation of Meinkoth and Wahl (1984) *Anal. Biochem.* 138:267-284:  $T_m = 81.5^\circ\text{C} + 16.6 (\log M) + 0.41 (\%GC) - 0.61 (\% \text{ form}) - 500/L$ ; where M is the molarity of monovalent cations, %GC is the percentage of guanosine and cytosine nucleotides in the DNA, % form is the percentage of formamide in the hybridization solution, and L is the length of the hybrid in base pairs. The  $T_m$  is the temperature (under defined ionic strength and pH) at which 50% of a complementary target sequence hybridizes to a perfectly matched probe.  $T_m$  is reduced by about 1°C for each 1% of mismatching; thus,  $T_m$ , hybridization, and/or wash conditions can be adjusted to hybridize to sequences of the desired identity. For example, if sequences with  $\geq 90\%$  identity are sought, the  $T_m$  can be decreased 10°C. Generally, stringent conditions are selected to be about 5°C lower than the thermal melting point ( $T_m$ ) for the specific sequence and its complement at a defined ionic strength and pH. However, severely stringent conditions can utilize a hybridization and/or wash at 1, 2, 3, or 4°C lower than the thermal melting point ( $T_m$ ); moderately stringent conditions can utilize a hybridization and/or wash at 6, 7, 8, 9, or 10°C lower than the thermal melting point ( $T_m$ ); low stringency conditions can utilize a hybridization and/or wash at 11, 12, 13, 14, 15, or 20°C lower than the thermal melting point ( $T_m$ ). Using the equation, hybridization and wash compositions, and desired  $T_m$ , those of ordinary skill will understand that variations in the stringency of hybridization and/or wash solutions are inherently described. If the desired degree of mismatching results in a  $T_m$  of less than 45°C (aqueous solution) or 32°C (formamide solution), it is preferred to increase the SSC concentration so that a higher temperature

can be used. An extensive guide to the hybridization of nucleic acids is found in Tijssen (1993) *Laboratory Techniques in Biochemistry and Molecular Biology—Hybridization with Nucleic Acid Probes*, Part I, Chapter 2 (Elsevier, New York); and Ausubel *et al.*, eds. (1995) *Current Protocols in Molecular Biology*, Chapter 2 (Greene Publishing and Wiley-Interscience, New York). See Sambrook *et al.* (1989) *Molecular Cloning: A Laboratory Manual* (2d ed., Cold Spring Harbor Laboratory Press, Plainview, New York).

Thus, isolated sequences that encode for a *Bt* toxin receptor protein and which hybridize under stringent conditions to the *Bt* toxin receptor sequences disclosed herein, or to fragments thereof, are encompassed by the present invention. Such sequences will be at least about 40% to 50% homologous, about 60%, 65%, or 70% homologous, and even at least about 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more homologous with the disclosed sequences. That is, the sequence identity of sequences may range, sharing at least about 40% to 50%, about 60%, 65%, or 70%, and even at least about 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more sequence identity.

The compositions and screening methods of the invention are useful for identifying cells expressing the BT toxin receptors of the invention, and variants and homologues thereof. Such identification could utilize detection methods at the protein level, such as ligand-receptor binding; or at the nucleotide level. Detection of the polypeptide could be *in situ* by means of *in situ* hybridization of tissue sections but may also be analyzed by bulk polypeptide purification and subsequent analysis by Western blot or immunological assay of a bulk preparation. Alternatively, receptor gene expression can be detected at the nucleic acid level by techniques well known to those of ordinary skill in any art using complimentary polynucleotides to assess the levels of genomic DNA, mRNA, and the like. As an example, PCR primers complimentary to the nucleic acid of interest can be used to identify the level of expression. Tissues and cells identified as expressing the receptor sequences of the invention are determined to be susceptible to toxins which bind the receptor polypeptides.

Where the source of the cells identified to express the receptor polypeptides of the invention is an organism, for example an insect plant pest, the organism is determined to be susceptible to toxins capable of binding the polypeptides. In a

particular embodiment, identification is in a lepidopteran plant pest expressing the *Bt* toxin receptor of the invention.

The invention encompasses antibody preparations with specificity against the polypeptides of the invention. In further embodiments of the invention, the antibodies  
5 are used to detect receptor expression in a cell.

In one aspect, the invention is particularly drawn to compositions and methods for modulating susceptibility of plant pests to *Bt* toxins. However, it is recognized that the methods and compositions could be used for modulating susceptibility of any cell or organism to the toxins. By "modulating" is intended that  
10 the susceptibility of a cell or organism to the cytotoxic effects of the toxin is increased or decreased. By "susceptibility" is intended that the viability of a cell contacted with the toxin is decreased. Thus the invention encompasses expressing the cell surface receptor polypeptides of the invention to increase susceptibility of a target cell or organ to *Bt* toxins. Such increases in toxin susceptibility are useful for medical and  
15 veterinary purposes in which eradication or reduction of viability of a group of cells is desired. Such increases in susceptibility are also useful for agricultural applications in which eradication or reduction of population of particular plant pests is desired.

Plant pests of interest include, but are not limited to insects, nematodes, and the like. Nematodes include parasitic nematodes such as root-knot, cyst, lesion, and  
20 reniform nematodes, etc.

The following examples are offered by way of illustration and not by way of limitation.

#### EXPERIMENTAL

##### 25 EXAMPLE 1: Isolation of EC *Bt* toxin receptor

Standard recombinant methods well known to those of ordinary skill in the art were carried out. For library construction, total RNA was isolated from the midgut of European corn borer (ECB), *Ostrinia nubilalis*. Corn borer larvae (for example, a mix of stage 2, 3, and 4, equal weight) can be pulverized in liquid nitrogen, homogenized, and  
30 total RNA extracted by standard procedures. PolyA RNA can be isolated from the total RNA with standard PolyA isolation procedures, such as the PolyATact system from Promega Corporation, Madison, WI. cDNA synthesis can then be performed and, for example, unidirectional cDNA libraries can be constructed according to known and

commercial procedures, such as the ZAP Express cDNA synthesis kit from Stratagene, La Jolla, CA. cDNA can be amplified by PCR, sized and properly digested with restriction fragments to be ligated into a vector. Subcloned cDNA can be sequenced to identify sequences with the proper peptide to identity corresponding to published sequences. These fragments can be used to probe genomic or cDNA libraries corresponding to a specific host, such as *Ostrinia nubilalis*, to obtain a full length coding sequence. Probes can also be made based on Applicants disclosed sequences. The coding sequence can then be ligated into a desired expression cassette and used to transform a host cell according to standard transformation procedures. Such an expression cassette can be part of a commercially available vector and expression system; for example, the pET system from Novagen Inc. (Madison, WI). Additional vectors that can be used for expression include pBKCMV, pBKRSV, pPbac and pMbac (Stratagene Inc.), pFASTBac1 (Gibco BRL) and other common bacterial, baculovirus, mammalian, and yeast expression vectors.

All vectors were constructed using standard molecular biology techniques as described for example in Sambrook *et al.*, (1989) *Molecular Cloning: A Laboratory Manual* (2<sup>nd</sup> ed., Cold Spring Harbor Laboratory: Cold Spring Harbor, N.Y.).

Expression is tested by ligand blotting and testing for *Bt* toxin binding. Ligand blotting, binding, and toxicity are tested by known methods; for example, as described in Martinez-Ramirez (1994) *Biochem. Biophys. Res. Comm.* 201: 782-787; Vadlamudi *et al.* (1995) *J Biol Chem* 270(10):5490-4, Keeton *et al.* (1998) *Appl Environ Microbiol* 64(6):2158-2165; Keeton *et al.* (1997) *Appl Environ Microbiol* 63(9):3419-3425; Ihara *et al.* (1998) *Comparative Biochemistry and Physiology, Part B* 120:197-204; Nagamatsu *et al.* (1998) *Biosci. Biotechnol. Biochem.* 62(4):718-726 and Nagamatsu *et al.* (1998) *Biosci. Biotechnol. Biochem.* 62(4):727-734.

Identifying the CryIA(b) binding polypeptide in ECB was done by ligand blotting brush border membrane vesicle polypeptides and probing those polypeptides for binding with CryIA(b) toxin. Two polypeptides, approximately 210 and 205 kDa, were found to bind to CryIA(b). Blotting and binding were done essentially as described in the preceding paragraph.

Degenerate primers for RT-PCR were designed based on known Cry1 toxin binding polypeptide sequences from *Manduca sexta* and *Bombyx mori*. The primers are shown below. cDNA was constructed from total midgut RNA (cDNA synthesis

kit GibcoBrL). Degenerate primers were used to amplify products of the expected size. The annealing temperature used was 53°C in generation of the 280 bp fragment and 55°C when generating the 1.6 kb fragment.

5 A 280bp fragment was obtained from ECB midgut RNA. Upon cloning and sequencing, the fragment was identified as having homology with the *Bt* toxin receptor 1 polypeptide (BTR1) described in Vadlamudi *et al.* (1995) *J Biol Chem* 270(10):5490-4.

A similar approach was used to generate a 1.6 kilobase pair clone. The sequence of primers used to generate the 280 base pair fragment were:  
10 Primer BTRD1S: 5'GTTAMYGTGAGAGAGGCAGAYCC3' (SEQ ID NO:8), and  
Primer BTRD5A: 5'GGATRTTAAGMGTCAGYACWCCG3' (SEQ ID NO:9).  
The sequence of primers used to generate the 1.6 kb fragment were:  
Primer BTRD6S: 5'TCCGAATTCTTCTTYAACCTCATCGAYAACTT3' (SEQ ID NO:10), and  
15 Primer BTRD7A: 5'CGCAAGCTTACTTGGTCGATGTTRCASGTCAT3' (SEQ ID NO:11)

The 1.6 kb fragment clone was ligated in an *E. coli* expression vector, pET-28a-c(+), and expressed using the pET system (Novagen Inc., Madison, WI). Purified polypeptide encoded by this 1.6kb fragment demonstrated binding to CryIA(b) in  
20 ligand blots. An ECB midgut cDNA library was generated and screened using this 1.6kb clone, generating 120 positive plaques. Thirty of these plaques were chosen for secondary screening and fifteen of those plaques were purified and sent for DNA sequencing.

The obtained nucleotide sequence of the selected *Bt* toxin receptor clone from  
25 ECB is set forth in SEQ ID NO: 1. The total length of the clone is 5498 base pairs. The coding sequences are residues 162-5312. The CryIA binding site is encoded by residues 4038-4547. The predicted transmembrane domain is encoded by residues 4872-4928. The corresponding deduced amino acid sequence for this *Bt* toxin receptor clone from ECB is set forth in SEQ ID NO: 2.

30 The purified polypeptide generated from the 1.6kb fragment set forth in SEQ ID NO:7 was used to inoculate rabbits for the production of polyclonal antibodies. On zoo western blots prepared from brush border membrane vesicles from various insect species, this set of antibodies specifically recognized ECB *Bt* toxin receptor

polypeptides, in comparison to *Bt* toxin receptor homologues polypeptides from other insect species. Rabbit polyclonal antibodies were also raised from a purified polypeptide corresponding to amino acids 1293-1462 of SEQ ID NO:2.

5 Example 2: Isolation of CEW and FAW *Bt* toxin receptor orthologues:

cDNA encoding a full-length *Bt* toxin receptor from corn earworm (CEW, *Heliothis Zea*) was isolated. The nucleotide sequence for this cDNA is set forth in SEQ ID NO: 3. Nucleotides 171-5360 correspond to the open reading frame. Nucleotides 4917-4973 correspond to the transmembrane region. Nucleotides 4083-4589 correspond to the CryIA binding site. The deduced corresponding amino acid sequence for the CEW *Bt* toxin receptor is set forth in SEQ ID NO: 4.

cDNA encoding a full-length *Bt* toxin receptor from fall armyworm (FAW, *Spodoptera frugiperda*) was isolated. The nucleotide sequence for this cDNA is set forth in SEQ ID NO: 5. Nucleotides 162-5363 correspond to the open reading frame. Nucleotides 4110-4616 correspond to the CryIA binding site. Nucleotides 4941-4997 correspond to the transmembrane region. Nucleotides 162-227 correspond to a signal peptide. The deduced corresponding amino acid sequence for the FAW *Bt* toxin receptor is set forth in SEQ ID NO: 6.

20 Example 3: Binding and cell death in *lepidopteran* insect cells expressing the *Bt* toxin receptors of the invention:

An *in vitro* system is developed to demonstrate the functionality of a *Bt* toxin receptor of the invention. The results disclosed in this example demonstrate that the ECB *Bt* toxin receptor of the invention (SEQ ID NOs:1 and 2) is specifically involved in the binding and killing action of CryIAb toxin.

Well known molecular biological methods are used in cloning and expressing the ECB *Bt* toxin receptor in Sf9 cells. A baculovirus expression system (Gibco BRL Catalogue No. 10359-016) is used according to the manufacturer's provided protocols and as described below. *S. frugiperda* (Sf9) cells obtained from ATCC (ATCC-CRL 1711) are grown at 27°C in Sf-900 II serum free medium (Gibco BRL, Catalogue No. 10902-088). These cells, which are not susceptible to CryIAb toxin, are transfected with an expression construct (pFastBac1 bacmid, Gibco BRL catalogue NO. 10360-014) comprising an operably linked *Bt* toxin receptor of the invention (SEQ ID NO:1)

downstream of a polyhedrin promoter. Transfected Sf9 cells express the ECB  $B_t$  toxin receptor and are lysed in the presence of Cry1Ab toxin. Toxin specificities, binding parameters, such as Kd values, and half maximal doses for cellular death and/or toxicity are also determined.

5 For generating expression constructs, the ECB  $B_t$  toxin receptor cDNA (SEQ ID NO:1) is subjected to appropriate restriction digestion, and the resulting cDNA comprising the full-length coding region is ligated into the donor plasmid pFastBac1 multiple cloning site. Following transformation and subsequent transposition, recombinant bacmid DNA comprising the ECB  $B_t$  toxin receptor (RBECB1) is  
10 isolated. As a control, recombinant bacmid DNA comprising the reporter gene  $\beta$ -glucuronidase (RBGUS) is similarly constructed and isolated.

For transfection, 2  $\mu$ g each RBECB1 or RBGUS DNA is mixed with 6  $\mu$ l of CellFectin (GibcoBRL catalogue NO. 10362-010) in 100  $\mu$ l of Sf900 medium, and incubated at room temperature for 30 minutes. The mixture is then diluted with 0.8 ml  
15 Sf-900 medium. Sf9 cells ( $10^6$ /ml per 35 mm well) are washed once with Sf-900 medium, mixed with the DNA/CellFectin mixture, added to the well, and incubated at room temperature for 5 hours. The medium is removed and 2 ml of Sf-900 medium containing penicillin and streptomycin is added to the well. 3-5 days after transfection, Western blotting is used to examine protein expression.

20 For Western blotting, 100  $\mu$ l of cell lysis buffer (50 mM Tris, pH7.8, 150mM NaCl, 1% Nonidet P-40) is added to the well. The cells are scraped and subjected to 16,000xg centrifugation. Pellet and supernatant are separated and subjected to Western blotting. An antibody preparation against ECB  $B_t$  toxin receptor (Example 1) is used as first antibody. Alkaline phosphatase-labelled anti-rabbit IgG is used as  
25 secondary antibody. Western blot results indicate that the full length ECB  $B_t$  toxin receptor of the invention (SEQ ID NOs:1 and 2) is expressed in the cell membrane of these cells.

For determining GUS activity, the medium of the cells transfected with RBGUS is removed. The cells and the medium are separately mixed with GUS  
30 substrate and assayed for the well known enzymatic activity. GUS activity assays indicate that this reporter gene is actively expressed in the transfected cells.

For determining toxin susceptibility, Cry toxins including but not limited to CryIA, CryIB, CryIC, CryID, CryIE, CryIF, CryII, Cry2, Cry3, and Cry9 toxins



(Schnepf E. *et al.* (1998) *Microbiology and Molecular Biology Reviews* 62(3): 775-806) are prepared by methods known in the art. Crystals are dissolved in pH 10.0, 50 mM carbonate buffer and treated with trypsin. Active fragments of Cry proteins are purified by chromatography. Three to five days after transfection, cells are washed  
5 with phosphate buffered saline (PBS). Different concentrations of active fragments of Cry toxins are applied to the cells. At different time intervals, the cells are examined under the microscope to readily determine susceptibility to the toxins. Alternatively, cell death, viability and/or toxicity is quantified by methods well known in the art. See, for example, In Situ Cell Death Detection Kits available from Roche  
10 Biochemicals (Catalogue Nos. 2 156 792, 1 684 809, and 1 684 817), and LIVE/DEAD® Viability/Cytotoxicity Kit available from Molecular Probes (catalogue No. L-3224).

A dose-dependent response of RBECB1-transfected cells to Cry1Ab is readily observed, with determined K<sub>d</sub> values well within the range for many receptors.  
15 Control cells, e.g. those transfected with pFastBac1 bacmid without an insert or those transfected with RBGus are not significantly affected by Cry1Ab. Interaction with other Cry toxins are similarly characterized.

This *in vitro* system is not only be used to verify the functionality of putative *Bt*-toxin receptors, but also used as a tool to determine the active site(s) and other  
20 functional domains of the toxin and the receptor. Furthermore, the system is used as a cell-based high throughput screen. For example, methods for distinguishing live versus dead cells by differential dyes are known in the art. This allows for aliquots of transfected cells to be treated with various toxin samples and to serve as a means for screening the toxin samples for desired specificity or binding characteristics. Since the  
25 system is used to identify the specificity of Cry protein receptors, it is a useful tool in insect resistance management.

Example 4: Expression of the ECB *Bt* toxin receptor in toxin susceptible stages of the insect's life cycle:

30 Total RNA was isolated from the eggs, pupae, adults, and the 1st through the 5th instar developmental stages, using TRIzol Reagent (Gibco BRL) essentially as instructed by the manufacturer.(Gibco BRL). The RNA was quantitated and 20 ug of each sample was loaded onto a formaldehyde agarose gel and electrophoresed at

constant voltage. The RNA was then transferred to a nylon membrane via neutral capillary transfer and cross-linked to the membrane using ultraviolet light. For hybridization, a 460 base pair ECB *Bt* toxin receptor DNA probe (bases 3682 to 4141 in SEQ ID NO:1) was constructed from a 460 base pair fragment prepared according to the manufacturer's protocol for Amersham Rediprime II random prime labeling system. The denatured probe was added to the membrane that had been prehybridized for at least 3 hours at 65°C and allowed to incubate with gentle agitation for at least 12 hours at 65°C. Following hybridization, the membranes were washed at 65°C for 1 hour with 1/4X 0.5M NaCl, 0.1M NaPO<sub>4</sub> (ph 7.0), 6mM EDTA and 1% SDS solution followed by two 1 hour washes in the above solution without SDS. The membrane was air dried briefly, wrapped in Saran Wrap and exposed to X-ray film.

An ECB *Bt* toxin receptor transcript of 5.5 kilobase was expressed strongly in the larval instars with much reduced expression in the pupal stage. The expression levels appeared to be fairly consistent from first to fifth instar, while decreasing markedly in the pupal stage. There were no detectable transcripts in either the egg or adult stages. These results indicate that the ECB *Bt* toxin transcript is being produced in the susceptible stages of the insects life cycle, while not being produced in stages resistant to the toxic effects of CryIAb.

#### Example 5: Tissue and subcellular expression of the ECB *Bt* toxin receptor:

Fifth instar ECB were dissected to isolate the following tissues: fat body (FB), malpighian tubules (MT), hind gut (HG), anterior midgut (AM) and posterior midgut (PM). Midguts from fifth instar larvae were also isolated for brush border membrane vesicle (BBMV) preparation using the well known protocol by Wolfersberger *et al.* (1987) *Comp. Biochem. Physiol.* 86A:301-308. Tissues were homogenized in Tris buffered saline, 0.1 % tween-20, centrifuged to pellet insoluble material, and transferred to a fresh tube. 50 ug of protein from each preparation was added to SDS sample buffer and B-mercaptoethanol, heated to 100°C for 10 minutes and loaded onto a 4-12% Bis-Tris gel (Novex). After electrophoresis, the proteins were transferred to a nitrocellulose membrane using a semi-dry apparatus. The membrane was blocked in 5% nonfat dry milk buffer for 1 hour at room temperature with gentle agitation. The primary antibody (Example 1) was added to a final dilution of 1:5000 and allowed to hybridize for 1 hour. The blot was then washed three times for 20

minutes each in nonfat milk buffer. The blot was then hybridized with the secondary antibody (goat anti-rabbit with alkaline phosphatase conjugate) at a dilution of 1:10000 for 1 hour at room temperature. Washes were performed as before. The bands were visualized by using the standard chemiluminescent protocol (Tropix western light protein detection kit).

The ECB *Bt* toxin receptor protein was only visible in the BBMV enriched lane, and not detected in any of the other ECB tissues types. This result indicates that the expression of the ECB *Bt* toxin receptor protein is at very low levels, since the BBMV preparation is a 20-30 fold enriched fraction of the midgut brush border. The result supports propositions that the ECB *Bt* toxin receptor is an integral membrane protein uniquely associated with the brush border. It also demonstrates that the ECB *Bt* toxin receptor is expressed in the envisioned target tissue for CryIAb toxins. However, the result does not necessarily rule out expression in other tissue types, albeit the expression of this protein in those tissues may be lower than in the BBMV enriched fraction.

All publications and patent applications mentioned in the specification are indicative of the level of those skilled in the art to which this invention pertains. All publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference.

Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be obvious that certain changes and modifications may be practiced within the scope of the appended claims.

Applicant's or agent's file reference	35718/204291	International application No.
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Date of deposit <b>11 July 2000 (11.07.00)</b>	Accession Number <b>PTA-2222</b>
<b>C. ADDITIONAL INDICATIONS</b> (leave blank if not applicable) This information is continued on an additional sheet <input type="checkbox"/>	
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## THAT WHICH IS CLAIMED:

1. An isolated nucleic acid molecule having a nucleotide sequence encoding a *Bt* toxin receptor, said sequence selected from the group consisting of:
  - a) a nucleotide sequence set forth in SEQ ID NO: 1, SEQ ID NO: 3 or SEQ ID NO: 5;
  - 5        b) a nucleotide sequence having at least about 60 % identity to the nucleotide sequence of a);
  - c) a nucleotide sequence having at least about 70 % identity to the nucleotide sequence of a);
  - d) a nucleotide sequence having at least about 75 % identity to the  
10        nucleotide sequence of a);
  - e) a nucleotide sequence having at least about 85 % identity to the nucleotide sequence of a);
  - f) a nucleotide sequence having at least about 95 % identity to the nucleotide sequence of a);
  - 15        g) a nucleotide sequence consisting of at least 22 contiguous nucleotides of the nucleotide sequence set forth in SEQ ID NO:1;
  - h) a nucleotide sequence consisting of at least about 15 contiguous nucleotides of the nucleotide sequence set forth in SEQ ID NO:3, or SEQ ID NO:5 ;
  - i) a nucleotide sequence that hybridizes under stringent  
20        conditions to the nucleotide sequence of a); and
2. The nucleic acid molecule of claim 1, wherein said toxin is a CryIA toxin.
- 25        3. The nucleic acid of claim 2, wherein said CryIA toxin is a CryIA(b) toxin.
4. An isolated polypeptide having the amino acid sequence selected from the group consisting of:
  - a) an amino acid sequence set forth in SEQ ID NO: 2, SEQ ID NO:  
30        4, or SEQ ID NO: 6;
  - b) an amino acid sequence having at least about 52% identity to

the amino acid sequence set forth in SEQ ID NO: 2;

c) an amino acid sequence having at least about 60 % identity to the amino acid sequence of a);

d) an amino acid sequence having at least about 70 % identity to the amino acid sequence of a);

e) an amino acid sequence having at least about 75 % identity to an amino acid sequence of a);

f) an amino acid sequence having at least about 85 % identity to an amino acid sequence of a);

g) an amino acid sequence having at least about 95 % identity to an amino acid sequence of a);

h) an amino acid comprising at least about 15 contiguous residues of the amino acid nucleotide sequence of a);

i) an amino acid sequence encoded by a nucleotide sequence according to claim 1;

j) a variant of the amino acid sequence of a);

k) a fragment of the amino acid sequence of a); and

l) a fragment of the amino acid sequence of a) that binds *Bt* toxin.

5. A fusion polypeptide comprising the polypeptide of claim 4, and at least one polypeptide of interest.

6. The fusion polypeptide of claim 5, wherein said polypeptide of interest is a toxin receptor.

7. An expression cassette comprising a nucleotide sequence encoding the fusion polypeptide of claim 5, wherein said nucleotide sequence is operably linked to a promoter that drives expression in a cell of interest.

8. The expression cassette of claim 7 wherein said polypeptide of interest is a toxin receptor.

9. An antibody preparation specific for the polypeptide of claim 4.



10. An expression cassette comprising at least one nucleotide sequence according to claim 1, wherein said nucleotide sequence is operably linked to a promoter that drives expression in a cell of interest.

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11. The expression cassette of claim 10, wherein said cell of interest is an insect or mammalian cell.

12. The expression cassette of claim 10 wherein said cell of interest is a  
10 microorganism.

13. The expression cassette of claim 12 wherein said microorganism is yeast or bacteria.

14. A vector for delivery of a nucleotide sequence to a cell of interest, the  
15 vector comprising at least one nucleotide sequence according to claim 1.

15. A cell containing the vector of claim 14.

16. A transformed cell of interest having stably incorporated within its  
20 genome a nucleotide sequence selected from the group consisting of:

a) a nucleotide sequence set forth in SEQ ID NO: 1, SEQ ID NO: 3  
or SEQ ID NO: 5;

b) a nucleotide sequence having at least about 60 % identity to the  
25 nucleotide sequence of a);

c) a nucleotide sequence having at least about 70 % identity to the  
nucleotide sequence of a);

d) a nucleotide sequence having at least about 75 % identity to the  
nucleotide sequence of a);

e) a nucleotide sequence having at least about 85 % identity to  
30 the nucleotide sequence of a);

f) a nucleotide sequence having at least about 95 % identity to the  
nucleotide sequence of a);

g) a nucleotide sequence consisting of at least 22 contiguous nucleotides of the nucleotide sequence set forth in SEQ ID NO:1;

h) a nucleotide sequence consisting of at least about 15 contiguous nucleotides of the nucleotide sequence set forth in SEQ ID NO:3, or SEQ ID NO:5 ;

i) a nucleotide sequence that hybridizes under stringent conditions to the nucleotide sequence of a); and

17. The transformed cell of claim 16 , wherein said cell is a plant cell.

10

18. The transformed cell of claim 17, wherein said plant cell is monocotyledonous.

19. A method for screening for ligands that bind *Bt* toxin receptor, said method comprising:

15

i) providing at least one *Bt* toxin receptor polypeptide according to claim 4;

ii) contacting said polypeptide with a sample and a control ligand under conditions promoting binding; and

20

iii) determining binding characteristics of said sample ligand, relative to said control ligand.

20. A method for screening for ligands that bind *Bt* toxin receptor, said method comprising:

25

i) providing at least one *Bt* toxin receptor polypeptide having the amino acid sequence selected from the group consisting of a, b, c, d, e, f, g, h , i, and j of claim 4 in cells expressing said polypeptide wherein said polypeptide comprises a toxin binding domain ;

ii) contacting said cells with a sample and a control ligand under conditions promoting binding; and

30

iii) determining binding characteristics of said sample ligand, relative to said control ligand.

21. The method of claim 20 wherein said toxin is a Cry1A toxin.

22. A method for screening for toxins that bind Bt toxin receptor, said method comprising the steps of claim 20; further comprising determining viability of said cells contacted with a sample ligand relative to said cells contacted with a control  
5 ligand.

23. The method of claim 20, wherein said sample ligand is a chimeric polypeptide comprising at least one primary polypeptide that binds a polypeptide having the amino acid sequence selected from the group consisting of:

10 a) an amino acid sequence set forth in SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 6;

b) an amino acid sequence having at least about 52% identity to the amino acid sequence set forth in SEQ ID NO: 2;

c) an amino acid sequence having at least about 60 % identity to the  
15 amino acid sequence of a);

d) an amino acid sequence having at least about 70 % identity to the amino acid sequence of a);

e) an amino acid sequence having at least about 75 % identity to an amino acid sequence of a);

20 f) an amino acid sequence having at least about 85 % identity to an amino acid sequence of a);

g) an amino acid sequence having at least about 95 % identity to an amino acid sequence of a);

h) an amino acid comprising at least about 15 contiguous residues of  
25 the amino acid nucleotide sequence of a);

i) an amino acid sequence encoded by a nucleotide sequence having at least about 60 % identity to the nucleotide sequence set forth in SEQ ID NO: 1, SEQ ID NO: 3 or SEQ ID NO: 5; and

j) a variant of the amino acid sequence of a).

30

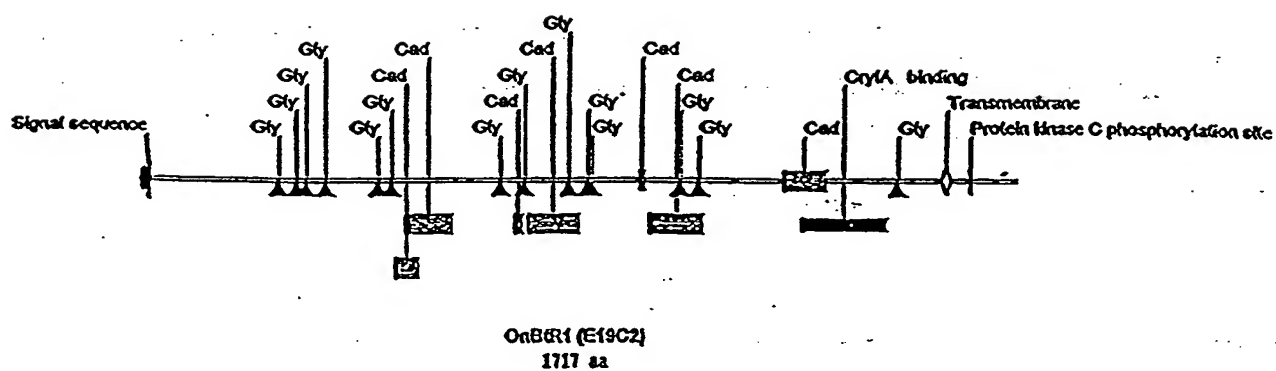
24. The method of claims 21, wherein said sample ligand is a chimeric polypeptide comprising at least one primary polypeptide that binds a polypeptide having the amino acid sequence selected from the group consisting of:

- a) an amino acid sequence set forth in SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 6;
- b) an amino acid sequence having at least about 52% identity to the amino acid sequence set forth in SEQ ID NO: 2;
- 5 c) an amino acid sequence having at least about 60 % identity to the amino acid sequence of a);
- d) an amino acid sequence having at least about 70 % identity to the amino acid sequence of a);
- e) an amino acid sequence having at least about 75 % identity to an amino acid sequence of a);
- 10 f) an amino acid sequence having at least about 85 % identity to an amino acid sequence of a);
- g) an amino acid sequence having at least about 95 % identity to an amino acid sequence of a);
- 15 h) an amino acid comprising at least about 15 contiguous residues of the amino acid nucleotide sequence of a);
- i) an amino acid sequence encoded by a nucleotide sequence having at least about 60 % identity to the nucleotide sequence set forth in SEQ ID NO: 1, SEQ ID NO: 3 or SEQ ID NO: 5; and
- 20 j) a variant of the amino acid sequence of a).

25. The method of claims 22, wherein said sample ligand is a chimeric polypeptide comprising at least one primary polypeptide that binds a polypeptide having the amino acid sequence selected from the group consisting of:

- a) an amino acid sequence set forth in SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 6;
- b) an amino acid sequence having at least about 52% identity to the amino acid sequence set forth in SEQ ID NO: 2;
- 30 c) an amino acid sequence having at least about 60 % identity to the amino acid sequence of a);
- d) an amino acid sequence having at least about 70 % identity to the amino acid sequence of a);

- e) an amino acid sequence having at least about 75 % identity to an amino acid sequence of a);
- f) an amino acid sequence having at least about 85 % identity to an amino acid sequence of a);
- 5 g) an amino acid sequence having at least about 95 % identity to an amino acid sequence of a);
- h) an amino acid comprising at least about 15 contiguous residues of the amino acid nucleotide sequence of a);
- i) an amino acid sequence encoded by a nucleotide sequence having at  
10 least about 60 % identity to the nucleotide sequence set forth in SEQ ID NO: 1, SEQ ID NO: 3 or SEQ ID NO: 5; and
- j) a variant of the amino acid sequence of a).



**Gly** = putative glycosilation sites

**Cad** = cadherin-like domain

**FIGURE 1**

## SEQUENCE LISTING

<110> Flannagan, Ronald D.  
Mathis, John P.  
Meyer, Terry E.

<120> Novel Bt Toxin Receptors From  
Lepidopteran Insects and Methods of Use

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Gln	Gln	Arg	Ser	Asn	Glu	Thr	Tyr	Thr	Leu	Val	Ile	Arg	Pro	Tyr	Asn		
1095						1100					1105						
ttc	cac	cac	cct	gtg	ttc	gtg	ttc	ccg	caa	ccc	gac	tcc	gtc	att	cgg	3536	
Phe	His	His	Pro	Val	Phe	Val	Phe	Pro	Gln	Pro	Asp	Ser	Val	Ile	Arg		
1110					1115					1120					1125		
ctt	tct	agg	gag	cgc	gca	aca	gaa	ggc	ggc	gtt	ctg	gcg	acg	gct	gcc	3584	
Leu	Ser	Arg	Glu	Arg	Ala	Thr	Glu	Gly	Gly	Val	Leu	Ala	Thr	Ala	Ala		
					1130					1135					1140		
aac	gag	ttc	ctg	gag	ccg	atc	tac	gcc	acc	gac	gag	gac	ggc	ctc	cac	3632	
Asn	Glu	Phe	Leu	Glu	Pro	Ile	Tyr	Ala	Thr	Asp	Glu	Asp	Gly	Leu	His		
			1145					1150					1155				
gcg	ggc	agc	gtc	acg	ttc	cac	gtc	cag	gga	aat	gag	gag	gcc	gtt	cag	3680	
Ala	Gly	Ser	Val	Thr	Phe	His	Val	Gln	Gly	Asn	Glu	Glu	Ala	Val	Gln		
			1160				1165						1170				
tac	ttt	gat	ata	act	gaa	gtg	gga	gca	gga	gaa	aat	agc	ggg	cag	ctt	3728	
Tyr	Phe	Asp	Ile	Thr	Glu	Val	Gly	Ala	Gly	Glu	Asn	Ser	Gly	Gln	Leu		
			1175			1180					1185						
ata	tta	cgc	cag	ctt	ttc	cca	gag	caa	atc	aga	caa	ttc	agg	atc	acg	3776	
Ile	Leu	Arg	Gln	Leu	Phe	Pro	Glu	Gln	Ile	Arg	Gln	Phe	Arg	Ile	Thr		
1190					1195					1200					1205		
atc	cgg	gcc	acg	gac	ggc	ggc	acg	gag	ccc	ggc	ccg	ctt	tgg	acc	gac	3824	
Ile	Arg	Ala	Thr	Asp	Gly	Gly	Thr	Glu	Pro	Gly	Pro	Leu	Trp	Thr	Asp		
				1210					1215					1220			
gtc	acg	ttt	tcg	gtg	gtc	ttc	gta	ccc	aca	cag	ggc	gac	cca	gtg	ttc	3872	
Val	Thr	Phe	Ser	Val	Val	Phe	Val	Pro	Thr	Gln	Gly	Asp	Pro	Val	Phe		
			1225					1230					1235				
agc	gaa	aat	gca	gct	act	gtc	gcc	ttc	ttc	gag	ggg	gaa	gaa	ggc	ctc	3920	
Ser	Glu	Asn	Ala	Ala	Thr	Val	Ala	Phe	Phe	Glu	Gly	Glu	Glu	Gly	Leu		
			1240				1245					1250					
cgt	gag	agt	ttt	gag	ctg	ccg	caa	gca	gaa	gac	ctt	aaa	aac	cac	ctc	3968	
Arg	Glu	Ser	Phe	Glu	Leu	Pro	Gln	Ala	Glu	Asp	Leu	Lys	Asn	His	Leu		
			1255			1260					1265						
tgc	gaa	gat	gac	tgc	caa	gat	atc	tac	tac	agg	ttt	att	gac	ggc	aac	4016	
Cys	Glu	Asp	Asp	Cys	Gln	Asp	Ile	Tyr	Tyr	Arg	Phe	Ile	Asp	Gly	Asn		
1270					1275					1280					1285		
aac	gag	ggg	ctt	ttc	gta	ctg	gac	cag	tca	agc	aac	gtc	atc	tcc	ctt	4064	
Asn	Glu	Gly	Leu	Phe	Val	Leu	Asp	Gln	Ser	Ser	Asn	Val	Ile	Ser	Leu		
				1290					1295					1300			
gcg	cag	gag	ttg	gac	cgc	gag	gtg	gcc	acg	tct	tac	acg	ctg	cac	atc	4112	
Ala	Gln	Glu	Leu	Asp	Arg	Glu	Val	Ala	Thr	Ser	Tyr	Thr	Leu	His	Ile		
			1305					1310					1315				
gcg	gcg	agc	aac	tcg	ccc	gac	gcc	act	ggg	atc	cct	ctg	cag	act	tcc	4160	
Ala	Ala	Ser	Asn	Ser	Pro	Asp	Ala	Thr	Gly	Ile	Pro	Leu	Gln	Thr	Ser		
			1320				1325				1330						
atc	ctc	gtt	gtc	acg	gtc	aat	gta	aga	gaa	gcg	aac	ccg	cgc	cca	att	4208	
Ile	Leu	Val	Val	Thr	Val	Asn	Val	Arg	Glu	Ala	Asn	Pro	Arg	Pro	Ile		
			1335			1340					1345						

ttc gag cag gac ctt tac aca gcg ggc att tcg acg ttg gac agc att Phe Glu Gln Asp Leu Thr Ala Gly Ile Ser Thr Leu Asp Ser Ile 1350 1355 1360 1365	4256
ggc cgg gaa ttg ctt act gtc agg gcg agc cac aca gaa gac gac acc Gly Arg Glu Leu Leu Thr Val Arg Ala Ser His Thr Glu Asp Asp Thr 1370 1375 1380	4304
atc acg tac acc ata gac cgt gcg agc atg cag ctg gac agc agc cta Ile Thr Tyr Thr Ile Asp Arg Ala Ser Met Gln Leu Asp Ser Ser Leu 1385 1390 1395	4352
gaa gcc gtg cgc gac tcg gcc ttc gcg ctg cat gcg acc acc ggc gtg Glu Ala Val Arg Asp Ser Ala Phe Ala Leu His Ala Thr Thr Gly Val 1400 1405 1410	4400
ctt tcg ctc aat atg cag ccc acc gct tcc atg cac ggc atg ttc gag Leu Ser Leu Asn Met Gln Pro Thr Ala Ser Met His Gly Met Phe Glu 1415 1420 1425	4448
ttc gac gtc atc gct acg gat aca gct tct gca atc gac aca gcc cgt Phe Asp Val Ile Ala Thr Asp Thr Ala Ser Ala Ile Asp Thr Ala Arg 1430 1435 1440 1445	4496
gtg aaa gtc tac ctc atc tca tcg caa aac cgc gtg acc ttc att ttc Val Lys Val Tyr Leu Ile Ser Ser Gln Asn Arg Val Thr Phe Ile Phe 1450 1455 1460	4544
gat aac caa ctt gag acc gtt gag cag aac aga aat ttc ata gcg gcc Asp Asn Gln Leu Glu Thr Val Glu Gln Asn Arg Asn Phe Ile Ala Ala 1465 1470 1475	4592
acg ttc agc acc ggg ttc aac atg acg tgc aac atc gac cag gtg gtg Thr Phe Ser Thr Gly Phe Asn Met Thr Cys Asn Ile Asp Gln Val Val 1480 1485 1490	4640
ccg ttc agc gac agc agc ggc gtg gcg caa gac gac acc acc gag gtg Pro Phe Ser Asp Ser Ser Gly Val Ala Gln Asp Asp Thr Thr Glu Val 1495 1500 1505	4688
cgc gcg cac ttc atc cgg gac aac gtg ccc gtg cag gca caa gag gtc Arg Ala His Phe Ile Arg Asp Asn Val Pro Val Gln Ala Gln Glu Val 1510 1515 1520 1525	4736
gag gcc gtc cgc agc gac acg gtg ctg ctg cgc acc atc cag ctg atg Glu Ala Val Arg Ser Asp Thr Val Leu Leu Arg Thr Ile Gln Leu Met 1530 1535 1540	4784
ctg agc acc aac agc ctg gtg ctg caa gac ctg gtg acg ggt gac act Leu Ser Thr Asn Ser Leu Val Leu Gln Asp Leu Val Thr Gly Asp Thr 1545 1550 1555	4832
ccg acg cta ggc gag gag tca atg cag atc gcc gtc tac gca cta gcc Pro Thr Leu Gly Glu Glu Ser Met Gln Ile Ala Val Tyr Ala Leu Ala 1560 1565 1570	4880
gcg ctc tcc gct gtg cta ggc ttc ctc tgc ctc gta ctg ctt ctc gca Ala Leu Ser Ala Val Leu Gly Phe Leu Cys Leu Val Leu Leu Leu Ala 1575 1580 1585	4928
ttg ttc tgt agg aca aga gca ctg aac cgg cag ctg caa gca ctc tcc	4976

Leu Phe Cys Arg Thr Arg Ala Leu Asn Arg Gln Leu Gln Ala Leu Ser  
 1590 1595 1600 1605  
 atg acg aag tac ggc tcg gtg gac tcc ggg ctg aac cgc gcc ggg ctg 5024  
 Met Thr Lys Tyr Gly Ser Val Asp Ser Gly Leu Asn Arg Ala Gly Leu  
 1610 1615 1620  
 gcg ccg ggc acc aac aag cac gcc gtc gag ggc tcc aac ccc atg tgg 5072  
 Ala Pro Gly Thr Asn Lys His Ala Val Glu Gly Ser Asn Pro Met Trp  
 1625 1630 1635  
 aac gag gcc atc cgc gcg ccc gac ttc gac gcc atc agt gac gcg agt 5120  
 Asn Glu Ala Ile Arg Ala Pro Asp Phe Asp Ala Ile Ser Asp Ala Ser  
 1640 1645 1650  
 ggc gac tcc gac ctg atc ggc atc gag gac atg ccg caa ttc cgc gac 5168  
 Gly Asp Ser Asp Leu Ile Gly Ile Glu Asp Met Pro Gln Phe Arg Asp  
 1655 1660 1665  
 gac tac ttc ccg ccc ggc gac aca gac tca agc agc ggc atc gtc ttg 5216  
 Asp Tyr Phe Pro Pro Gly Asp Thr Asp Ser Ser Ser Gly Ile Val Leu  
 1670 1675 1680 1685  
 cac atg ggc gaa gcc acg gac aac aag ccc gtg acc acg cat ggc aac 5264  
 His Met Gly Glu Ala Thr Asp Asn Lys Pro Val Thr Thr His Gly Asn  
 1690 1695 1700  
 aac ttc ggg ttc aag tcc acc ccg tac ctg cca cag ccg cac cca aag 5312  
 Asn Phe Gly Phe Lys Ser Thr Pro Tyr Leu Pro Gln Pro His Pro Lys  
 1705 1710 1715  
 taactgccag ggtataacct gtccagggtg cctacgccgc gcgaagtgcg cacacgcgtt 5372  
 tatcatcggg aaacattagc atgaagatac ctatgtacat attgtaaaatt gtaacatatac 5432  
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 ctcgag 5498

&lt;210&gt; 2

&lt;211&gt; 1717

&lt;212&gt; PRT

<213> *Ostrinia nubilalis*

&lt;400&gt; 2

Met Gly Val Glu Arg Phe Phe Pro Ala Val Leu Leu Val Ser Leu Ala  
 1 5 10 15  
 Ser Ala Ala Leu Ala Asn Gln Arg Cys Ser Tyr Ile Ile Ala Ile Pro  
 20 25 30  
 Arg Pro Glu Thr Pro Glu Leu Pro Pro Ile Asp Tyr Glu Gly Lys Ser  
 35 40 45  
 Trp Ser Glu Gln Pro Leu Ile Pro Gly Pro Thr Arg Glu Glu Val Cys  
 50 55 60  
 Met Glu Asn Phe Leu Pro Asp Gln Met Ile Gln Val Ile Tyr Met Glu  
 65 70 75 80  
 Glu Glu Ile Glu Gly Asp Val Ile Ile Ala Lys Leu Asn Tyr Gln Gly  
 85 90 95  
 Ser Asn Thr Pro Val Leu Ser Ile Met Ser Gly Gln Pro Arg Ala Gln  
 100 105 110  
 Leu Gly Pro Glu Phe Arg Gln Asn Glu Ala Asp Gly Gln Trp Ser Leu  
 115 120 125  
 Val Ile Thr Gln Arg Gln Asp Tyr Glu Thr Ala Thr Met Gln Ser Tyr  
 130 135 140  
 Val Phe Ser Ile Gln Val Glu Gly Glu Ser Gln Ala Val Leu Val Ala  
 145 150 155 160

Leu Glu Ile Val Asn Ile Asp Asp Asn Pro Pro Ile Leu Gln Val Val  
 165 170 175  
 Ser Ala Cys Val Ile Pro Glu His Gly Glu Ala Arg Leu Thr Asp Cys  
 180 185 190  
 Val Tyr Gln Val Ser Asp Arg Asp Gly Glu Ile Ser Thr Arg Phe Met  
 195 200 205  
 Thr Phe Arg Val Asp Ser Ser Arg Ala Ala Asp Glu Ser Ile Phe Tyr  
 210 215 220  
 Met Val Gly Glu Tyr Asp Pro Ser Asp Trp Phe Asn Met Lys Met Thr  
 225 230 235 240  
 Val Gly Ile Asn Ser Pro Leu Asn Phe Glu Thr Thr Gln Leu His Ile  
 245 250 255  
 Phe Ser Val Thr Ala Ser Asp Ser Leu Pro Asn Asn His Thr Val Thr  
 260 265 270  
 Met Met Val Gln Val Glu Asn Val Glu Ser Arg Pro Pro Arg Trp Val  
 275 280 285  
 Glu Ile Phe Ser Val Gln Gln Phe Asp Glu Lys Thr Asn Gln Ser Phe  
 290 295 300  
 Ser Leu Arg Ala Ile Asp Gly Asp Thr Gly Ile Asn Arg Ala Ile Asn  
 305 310 315 320  
 Tyr Thr Leu Ile Arg Asp Asp Ala Asp Asp Phe Phe Ser Leu Glu Val  
 325 330 335  
 Ile Glu Asp Gly Ala Ile Leu His Val Thr Glu Ile Asp Arg Asp Lys  
 340 345 350  
 Leu Glu Arg Glu Leu Phe Asn Leu Thr Ile Val Ala Tyr Lys Ser Thr  
 355 360 365  
 Asp Ala Ser Phe Ala Thr Glu Ala His Ile Phe Ile Ile Val Asn Asp  
 370 375 380  
 Val Asn Asp Gln Arg Pro Glu Pro Leu His Lys Glu Tyr Ser Ile Asp  
 385 390 395 400  
 Ile Met Glu Glu Thr Pro Met Thr Leu Asn Phe Asn Glu Glu Phe Gly  
 405 410 415  
 Phe His Asp Arg Asp Leu Gly Glu Asn Ala Gln Tyr Thr Val Glu Leu  
 420 425 430  
 Glu Asp Val Phe Pro Pro Gly Ala Ala Ser Ala Phe Tyr Ile Ala Pro  
 435 440 445  
 Gly Ser Gly Tyr Gln Arg Gln Thr Phe Ile Met Gly Thr Ile Asn His  
 450 455 460  
 Thr Met Leu Asp Tyr Glu Asp Val Ile Phe Gln Asn Ile Ile Ile Lys  
 465 470 475 480  
 Val Lys Ala Val Asp Met Asn Asn Ala Ser His Val Gly Glu Ala Leu  
 485 490 495  
 Val Tyr Val Asn Leu Ile Asn Trp Asn Asp Glu Leu Pro Ile Phe Glu  
 500 505 510  
 Glu Ser Ser Tyr Ser Ala Ser Phe Lys Glu Thr Val Gly Ala Gly Phe  
 515 520 525  
 Pro Val Ala Thr Val Leu Ala Leu Asp Arg Asp Ile Asp Asp Val Val  
 530 535 540  
 Val His Ser Leu Met Gly Asn Ala Val Asp Tyr Leu Phe Ile Asp Glu  
 545 550 555 560  
 Ser Thr Gly Glu Ile Phe Val Ser Met Asp Asp Ala Phe Asp Tyr His  
 565 570 575  
 Arg Gln Asn Thr Leu Phe Val Gln Val Arg Ala Asp Asp Thr Leu Gly  
 580 585 590  
 Asp Gly Pro His Asn Thr Val Thr Thr Gln Leu Val Ile Glu Leu Glu  
 595 600 605  
 Asp Val Asn Asn Thr Pro Pro Thr Leu Arg Leu Pro Arg Ser Thr Pro  
 610 615 620  
 Ser Val Glu Glu Asn Val Pro Glu Gly Tyr Glu Ile Ser Arg Glu Ile  
 625 630 635 640  
 Thr Ala Thr Asp Pro Asp Thr Ser Ala Tyr Leu Trp Phe Glu Ile Asp  
 645 650 655

Trp Asp Ser Thr Trp Ala Thr Lys Gln Gly Arg Glu Thr Asn Pro Thr  
 660 665 670  
 Glu Tyr Val Gly Cys Ile Val Ile Glu Thr Ile Tyr Pro Thr Glu Gly  
 675 680 685  
 Asn Arg Gly Ser Ala Ile Gly Arg Leu Val Val Gln Glu Ile Arg Asp  
 690 695 700  
 Asn Val Thr Ile Asp Phe Glu Glu Phe Glu Met Leu Tyr Leu Thr Val  
 705 710 715 720  
 Arg Val Arg Asp Leu Asn Thr Val Ile Gly Asp Asp Tyr Asp Glu Ala  
 725 730 735  
 Thr Phe Thr Ile Thr Ile Ile Asp Met Asn Asp Asn Ala Pro Ile Phe  
 740 745 750  
 Ala Asn Gly Thr Leu Thr Gln Thr Met Arg Val Arg Glu Leu Ala Ala  
 755 760 765  
 Ser Gly Thr Leu Ile Gly Ser Val Leu Ala Thr Asp Ile Asp Gly Pro  
 770 775 780  
 Leu Tyr Asn Gln Val Arg Tyr Thr Ile Gln Pro Arg Asn Asn Thr Pro  
 785 790 795 800  
 Glu Gly Leu Val Lys Ile Asp Phe Thr Thr Gly Gln Ile Glu Val Asp  
 805 810 815  
 Ala Asn Glu Ala Ile Asp Ala Asp Glu Pro Trp Arg Phe Tyr Leu Tyr  
 820 825 830  
 Tyr Thr Val Ile Ala Ser Asp Glu Cys Ser Leu Glu Asn Arg Thr Glu  
 835 840 845  
 Cys Pro Pro Asp Ser Asn Tyr Phe Glu Val Pro Gly Asp Ile Glu Ile  
 850 855 860  
 Glu Ile Ile Asp Thr Asn Asn Lys Val Pro Glu Pro Leu Thr Glu Lys  
 865 870 875 880  
 Phe Asn Thr Thr Val Tyr Val Trp Glu Asn Ala Thr Ser Gly Asp Glu  
 885 890 895  
 Val Val Gln Leu Tyr Ser His Asp Arg Asp Arg Asp Glu Leu Tyr His  
 900 905 910  
 Thr Val Arg Tyr Thr Met Asn Phe Ala Val Asn Pro Arg Leu Arg Asp  
 915 920 925  
 Phe Phe Glu Val Asp Leu Asp Thr Gly Arg Leu Glu Val His Tyr Pro  
 930 935 940  
 Gly Asp Glu Lys Leu Asp Arg Asp Gly Asp Glu Pro Thr His Thr Ile  
 945 950 955 960  
 Phe Val Asn Phe Ile Asp Asn Phe Phe Ser Asp Gly Asp Gly Arg Arg  
 965 970 975  
 Asn Gln Asp Glu Val Glu Ile Phe Val Val Leu Leu Asp Val Asn Asp  
 980 985 990  
 Asn Ala Pro Glu Met Pro Leu Pro Asp Glu Leu Arg Phe Asp Val Ser  
 995 1000 1005  
 Glu Gly Ala Val Ala Gly Val Arg Val Leu Pro Glu Ile Tyr Ala Pro  
 1010 1015 1020  
 Asp Arg Asp Glu Pro Asp Thr Asp Asn Ser Arg Val Gly Tyr Gly Ile  
 1025 1030 1035 1040  
 Leu Asp Leu Thr Ile Thr Asp Arg Asp Ile Glu Val Pro Asp Leu Phe  
 1045 1050 1055  
 Thr Met Ile Ser Ile Glu Asn Lys Thr Gly Glu Leu Glu Thr Ala Met  
 1060 1065 1070  
 Asp Leu Arg Gly Tyr Trp Gly Thr Tyr Glu Ile Phe Ile Glu Ala Phe  
 1075 1080 1085  
 Asp His Gly Tyr Pro Gln Gln Arg Ser Asn Glu Thr Tyr Thr Leu Val  
 1090 1095 1100  
 Ile Arg Pro Tyr Asn Phe His His Pro Val Phe Val Phe Pro Gln Pro  
 1105 1110 1115 1120  
 Asp Ser Val Ile Arg Leu Ser Arg Glu Arg Ala Thr Glu Gly Gly Val  
 1125 1130 1135  
 Leu Ala Thr Ala Ala Asn Glu Phe Leu Glu Pro Ile Tyr Ala Thr Asp  
 1140 1145 1150



Glu Asp Gly Leu His Ala Gly Ser Val Thr Phe His Val Gln Gly Asn  
 1155 1160 1165  
 Glu Glu Ala Val Gln Tyr Phe Asp Ile Thr Glu Val Gly Ala Gly Glu  
 1170 1175 1180  
 Asn Ser Gly Gln Leu Ile Leu Arg Gln Leu Phe Pro Glu Gln Ile Arg  
 1185 1190 1195 1200  
 Gln Phe Arg Ile Thr Ile Arg Ala Thr Asp Gly Gly Thr Glu Pro Gly  
 1205 1210 1215  
 Pro Leu Trp Thr Asp Val Thr Phe Ser Val Val Phe Val Pro Thr Gln  
 1220 1225 1230  
 Gly Asp Pro Val Phe Ser Glu Asn Ala Ala Thr Val Ala Phe Phe Glu  
 1235 1240 1245  
 Gly Glu Glu Gly Leu Arg Glu Ser Phe Glu Leu Pro Gln Ala Glu Asp  
 1250 1255 1260  
 Leu Lys Asn His Leu Cys Glu Asp Asp Cys Gln Asp Ile Tyr Tyr Arg  
 1265 1270 1275 1280  
 Phe Ile Asp Gly Asn Asn Glu Gly Leu Phe Val Leu Asp Gln Ser Ser  
 1285 1290 1295  
 Asn Val Ile Ser Leu Ala Gln Glu Leu Asp Arg Glu Val Ala Thr Ser  
 1300 1305 1310  
 Tyr Thr Leu His Ile Ala Ala Ser Asn Ser Pro Asp Ala Thr Gly Ile  
 1315 1320 1325  
 Pro Leu Gln Thr Ser Ile Leu Val Val Thr Val Asn Val Arg Glu Ala  
 1330 1335 1340  
 Asn Pro Arg Pro Ile Phe Glu Gln Asp Leu Tyr Thr Ala Gly Ile Ser  
 1345 1350 1355 1360  
 Thr Leu Asp Ser Ile Gly Arg Glu Leu Leu Thr Val Arg Ala Ser His  
 1365 1370 1375  
 Thr Glu Asp Asp Thr Ile Thr Tyr Thr Ile Asp Arg Ala Ser Met Gln  
 1380 1385 1390  
 Leu Asp Ser Ser Leu Glu Ala Val Arg Asp Ser Ala Phe Ala Leu His  
 1395 1400 1405  
 Ala Thr Thr Gly Val Leu Ser Leu Asn Met Gln Pro Thr Ala Ser Met  
 1410 1415 1420  
 His Gly Met Phe Glu Phe Asp Val Ile Ala Thr Asp Thr Ala Ser Ala  
 1425 1430 1435 1440  
 Ile Asp Thr Ala Arg Val Lys Val Tyr Leu Ile Ser Ser Gln Asn Arg  
 1445 1450 1455  
 Val Thr Phe Ile Phe Asp Asn Gln Leu Glu Thr Val Glu Gln Asn Arg  
 1460 1465 1470  
 Asn Phe Ile Ala Ala Thr Phe Ser Thr Gly Phe Asn Met Thr Cys Asn  
 1475 1480 1485  
 Ile Asp Gln Val Val Pro Phe Ser Asp Ser Ser Gly Val Ala Gln Asp  
 1490 1495 1500  
 Asp Thr Thr Glu Val Arg Ala His Phe Ile Arg Asp Asn Val Pro Val  
 1505 1510 1515 1520  
 Gln Ala Gln Glu Val Glu Ala Val Arg Ser Asp Thr Val Leu Leu Arg  
 1525 1530 1535  
 Thr Ile Gln Leu Met Leu Ser Thr Asn Ser Leu Val Leu Gln Asp Leu  
 1540 1545 1550  
 Val Thr Gly Asp Thr Pro Thr Leu Gly Glu Glu Ser Met Gln Ile Ala  
 1555 1560 1565  
 Val Tyr Ala Leu Ala Ala Leu Ser Ala Val Leu Gly Phe Leu Cys Leu  
 1570 1575 1580  
 Val Leu Leu Leu Ala Leu Phe Cys Arg Thr Arg Ala Leu Asn Arg Gln  
 1585 1590 1595 1600  
 Leu Gln Ala Leu Ser Met Thr Lys Tyr Gly Ser Val Asp Ser Gly Leu  
 1605 1610 1615  
 Asn Arg Ala Gly Leu Ala Pro Gly Thr Asn Lys His Ala Val Glu Gly  
 1620 1625 1630  
 Ser Asn Pro Met Trp Asn Glu Ala Ile Arg Ala Pro Asp Phe Asp Ala  
 1635 1640 1645

Ile Ser Asp Ala Ser Gly Asp Ser Asp Leu Ile Gly Ile Glu Asp Met  
 1650 1655 1660  
 Pro Gln Phe Arg Asp Asp Tyr Phe Pro Pro Gly Asp Thr Asp Ser Ser  
 1665 1670 1675 1680  
 Ser Gly Ile Val Leu His Met Gly Glu Ala Thr Asp Asn Lys Pro Val  
 1685 1690 1695  
 Thr Thr His Gly Asn Asn Phe Gly Phe Lys Ser Thr Pro Tyr Leu Pro  
 1700 1705 1710  
 Gln Pro His Pro Lys  
 1715

&lt;210&gt; 3

&lt;211&gt; 5527

&lt;212&gt; DNA

&lt;213&gt; Heliothis zea

&lt;220&gt;

&lt;221&gt; CDS

&lt;222&gt; (171)... (5360)

&lt;400&gt; 3

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 atttgtgtaa agtgtagtgt taaataattt ggcattgctg taaaggatta aagagtgttc 120  
 caattgatca cccagagggt gatcgaccag actagacaca gaactatgag atg gca 176  
 Met Ala  
 1

gtc gac gtg aga ata ttg acg gca gcg gtt ttc att atc gct gct cac 224  
 Val Asp Val Arg Ile Leu Thr Ala Ala Val Phe Ile Ile Ala Ala His  
 5 10 15

ttg act ttc gcg caa gat tgt agc tac atg gta gca ata ccc aga cca 272  
 Leu Thr Phe Ala Gln Asp Cys Ser Tyr Met Val Ala Ile Pro Arg Pro  
 20 25 30

gag cga cca gat ttt cca agt cta aat ttc gat gga ata cca tgg agt 320  
 Glu Arg Pro Asp Phe Pro Ser Leu Asn Phe Asp Gly Ile Pro Trp Ser  
 35 40 45 50

cgg tat ccc ctg ata cca gtg gag ggt aga gaa gat gtg tgc atg aac 368  
 Arg Tyr Pro Leu Ile Pro Val Glu Gly Arg Glu Asp Val Cys Met Asn  
 55 60 65

gaa ttc cag cca gat gcc ttg aac cca gtt acc gtc atc ttc atg gag 416  
 Glu Phe Gln Pro Asp Ala Leu Asn Pro Val Thr Val Ile Phe Met Glu  
 70 75 80

gag gag ata gaa ggg gat gtg gct atc gcg agg ctt aac tac cga ggt 464  
 Glu Glu Ile Glu Gly Asp Val Ala Ile Ala Arg Leu Asn Tyr Arg Gly  
 85 90 95

acc aat act ccg acc att gta tct cca ttt agc ttt ggt act ttt aac 512  
 Thr Asn Thr Pro Thr Ile Val Ser Pro Phe Ser Phe Gly Thr Phe Asn  
 100 105 110

atg ttg ggg ccg gtc ata cgt aga ata cct gag aat ggt ggc gac tgg 560  
 Met Leu Gly Pro Val Ile Arg Arg Ile Pro Glu Asn Gly Gly Asp Trp  
 115 120 125 130

cat ctc gtc att aca cag aga cag gac tac gag acg cca ggt atg cag 608  
 His Leu Val Ile Thr Gln Arg Gln Asp Tyr Glu Thr Pro Gly Met Gln  
 135 140 145

cag tac atc ttc gac gtg agg gta gac gat gaa ccg cta gtg gcc acg Gln Tyr Ile Phe Asp Val Arg Val Asp Asp Glu Pro Leu Val Ala Thr 150 155 160	-656
gtg atg ctg ctc att gtc aac atc gat gac aac gat cct atc ata cag Val Met Leu Leu Ile Val Asn Ile Asp Asp Asn Asp Pro Ile Ile Gln 165 170 175	704
atg ttt gag cct tgt gat att cct gaa cgc ggt gaa aca ggc atc aca Met Phe Glu Pro Cys Asp Ile Pro Glu Arg Gly Glu Thr Gly Ile Thr 180 185 190	752
tca tgc aag tac acc gtg agc gat gct gac ggc gag atc agt aca cgt Ser Cys Lys Tyr Thr Val Ser Asp Ala Asp Gly Glu Ile Ser Thr Arg 195 200 205 210	800
ttc atg agg ttc gaa atc agc agc gat cga gac gat gac gaa tat ttc Phe Met Arg Phe Glu Ile Ser Ser Asp Asp Asp Asp Glu Tyr Phe 215 220 225	848
gaa ctc gtc aga gaa aat ata caa gga caa tgg atg tat gtt cat atg Glu Leu Val Arg Glu Asn Ile Gln Gly Gln Trp Met Tyr Val His Met 230 235 240	896
aga gtt cac gtc aaa aaa cct ctt gat tat gag gaa aac ccg cta cat Arg Val His Val Lys Lys Pro Leu Asp Tyr Glu Glu Asn Pro Leu His 245 250 255	944
ttg ttt aga gtt aca gct tat gat tcc cta cca aac aca cat aca gtg Leu Phe Arg Val Thr Ala Tyr Asp Ser Leu Pro Asn Thr His Thr Val 260 265 270	992
acg atg atg gtg caa gta gag aac gtt gag aac aga ccg ccg cga tgg Thr Met Met Val Gln Val Glu Asn Val Glu Asn Arg Pro Pro Arg Trp 275 280 285 290	1040
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caa acg ata gaa ggt ggt cga gaa ggc gct tgg ttt aac gtc gct cca Gln Thr Ile Glu Gly Gly Arg Glu Gly Ala Trp Phe Asn Val Ala Pro 340 345 350	1232
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tcg aaa acc gat gtg gtc atc atc gtg aac gat gtc aat gat cag gcg	1376

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			390					395					400			
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Pro	Leu	Pro	Phe	Arg	Glu	Glu	Tyr	Ser	Ile	Glu	Ile	Met	Glu	Glu	Thr	
		405					410					415				
gcg	atg	acc	ctg	aat	tta	gaa	gac	ttt	ggg	ttc	cat	gat	aga	gat	ctc	1472
Ala	Met	Thr	Leu	Asn	Leu	Glu	Asp	Phe	Gly	Phe	His	Asp	Arg	Asp	Leu	
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ggt	cct	cac	gca	caa	tac	aca	gta	cac	tta	gag	agc	atc	cat	cct	ccc	1520
Gly	Pro	His	Ala	Gln	Tyr	Thr	Val	His	Leu	Glu	Ser	Ile	His	Pro	Pro	
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Arg	Ala	His	Glu	Ala	Phe	Tyr	Ile	Ala	Pro	Glu	Val	Gly	Tyr	Gln	Arg	
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cag	tcc	ttc	att	atg	ggc	acg	cag	aac	cat	cac	atg	ctg	gac	ttc	gaa	1616
Gln	Ser	Phe	Ile	Met	Gly	Thr	Gln	Asn	His	His	Met	Leu	Asp	Phe	Glu	
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Val	Pro	Glu	Phe	Gln	Asn	Ile	Gln	Leu	Arg	Ala	Val	Gln	Ala	Ile	Asp	
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Asp	Asp	Pro	Lys	Trp	Val		Ile	Ala	Ile	Ile	Asn	Ile	Lys	Leu	Ile	
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Asn	Trp	Asn	Asp	Glu	Leu	Pro	Met	Phe	Glu	Ser	Asp	Val	Gln	Thr	Val	
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Ser	Phe	Asp	Glu	Thr	Glu	Gly	Ala	Gly	Phe	Tyr	Val	Ala	Thr	Val	Val	
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gcg	aag	gac	cgg	gat	gtt	ggt	gat	aaa	gtc	gaa	cac	tct	cta	atg	ggt	1856
Ala	Lys	Asp	Arg	Asp	Val	Gly	Asp	Lys	Val	Glu	His	Ser	Leu	Met	Gly	
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Asn	Ala	Val	Ser	Tyr	Leu	Arg	Ile	Asp	Lys	Glu	Thr	Gly	Glu	Ile	Phe	
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Val	Thr	Glu	Asn	Glu	Ala	Phe	Asn	Tyr	His	Arg	Gln	Asn	Glu	Leu	Phe	
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acg	ctc	agg	ctg	cct	cg	gcc	act	cca	tca	gtg	gaa	gag	aac	gtg	ccc	2096
Thr	Leu	Arg	Leu	Pro	Arg	Ala	Thr	Pro	Ser	Val	Glu	Glu	Asn	Val	Pro	
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atc gag acg ata tac ccg aat ata aac cag cga ggc aac gcc atc ggc Ile Glu Thr Ile Tyr Pro Asn Ile Asn Gln Arg Gly Asn Ala Ile Gly 695 700 705	2288
cgc gtg gta gtg cga gag atc cgg gac ggc gtc acc ata gac tat gag Arg Val Val Val Arg Glu Ile Arg Asp Gly Val Thr Ile Asp Tyr Glu 710 715 720	2336
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gtt att gga gaa gac cat gat ata tcc aca ttc acg atc acg ata ata Val Ile Gly Glu Asp His Asp Ile Ser Thr Phe Thr Ile Thr Ile Ile 740 745 750	2432
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Lys	Val	Pro	Gln	Val	Glu	Asp	Asp	Lys	Phe	Glu	Ala	Thr	Val	Tyr	Ile		
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Tyr	Glu	Gly	Ala	Asp	Asp	Gly	Gln	His	Val	Val	Gln	Ile	Tyr	Ala	Ser		
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Pro	Gln	Arg	Ile	Ser	Asn	Gln	Lys	Tyr	Pro	Leu	Val	Ile	Arg	Pro	Tyr		
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 Met Glu Glu Glu Ile Glu Gly Asp Val Ala Ile Ala Arg Leu Asn Tyr  
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 Phe Asn Met Leu Gly Pro Val Ile Arg Arg Ile Pro Glu Asn Gly Gly  
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Leu His Leu Phe Arg	Val Thr Ala Tyr Asp	Ser Leu Pro Asn Thr His			
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Thr Val Thr Met Met	Val Gln Val Glu Asn Val	Glu Asn Arg Pro Pro			
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Arg Trp Met Glu Ile	Phe Ala Val Gln Gln Phe	Asp Glu Lys Thr Glu			
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Phe Gln Ser Lys Thr	Asp Val Val Ile Ile	Val Asn Asp Val Asn Asp			
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Glu Thr Ala Met Thr	Leu Asn Leu Glu Asp	Phe Gly Phe His Asp Arg			
	420	425	430		
Asp Leu Gly Pro His	Ala Gln Tyr Thr Val	His Leu Glu Ser Ile His			
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Pro Pro Arg Ala His	Glu Ala Phe Tyr Ile	Ala Pro Glu Val Gly Tyr			
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Asp Met Asp Asp Pro	Lys Trp Val Gly Ile	Ala Ile Ile Asn Ile Lys			
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Met Gly Asn Ala Val	Ser Tyr Leu Arg Ile	Asp Lys Glu Thr Gly Glu			
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Pro Pro Thr Leu Arg	Leu Pro Arg Ala Thr	Pro Ser Val Glu Glu Asn			
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Ala Thr Lys Gln Gly	Arg Asn Thr Asp	Ser Lys Glu Tyr Ile Gly Cys			
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# INTERNATIONAL SEARCH REPORT

Intern. Appl. No.

PCT/US 00/31674

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C12N15/12 C12N15/62 C12N5/10 C07K14/705 C07K16/28  
G01N33/50

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12N C07K G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

WPI Data, PAJ, CAB Data, STRAND, BIOSIS, EPO-Internal

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	NAGAMATSU YASUNORI ET AL: "Cloning, sequencing, and expression of the Bombyx mori receptor for Bacillus thuringiensis insecticidal CryIA(a) toxin." BIOSCIENCE BIOTECHNOLOGY AND BIOCHEMISTRY, vol. 62, no. 4, April 1998 (1998-04), pages 727-734, XP002164759 ISSN: 0916-8451 cited in the application the whole document	1,2,4, 9-15
X	WO 96 12964 A (UNIV WYOMING) 2 May 1996 (1996-05-02) the whole document	1,2,4, 9-15, 19-22

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

### \* Special categories of cited documents:

- \*A\* document defining the general state of the art which is not considered to be of particular relevance
- \*E\* earlier document but published on or after the international filing date
- \*L\* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- \*O\* document referring to an oral disclosure, use, exhibition or other means
- \*P\* document published prior to the international filing date but later than the priority date claimed

- \*T\* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- \*X\* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- \*Y\* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- \*S\* document member of the same patent family

Date of the actual completion of the international search

13 June 2001

Date of mailing of the international search report

29.06.01

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2  
NL - 2280 HV Rijswijk  
Tel: (+31-70) 340-2040; Tx: 31 651 epo nl;  
Fax: (+31-70) 340-3016

Authorized officer

Hornig, H

# INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 00/31674

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	VADLAMUDI RATNA K ET AL: "Cloning and expression of a receptor for an insecticidal toxin of <i>Bacillus thuringiensis</i> ." JOURNAL OF BIOLOGICAL CHEMISTRY, vol. 270, no. 10, 1995, pages 5490-5494; XP002164760 ISSN: 0021-9258 cited in the application the whole document	1,2,4, 9-15
X	KEETON TIMOTHY P ET AL: "Ligand specificity and affinity of BT-R-1, the <i>Bacillus thuringiensis</i> CryIA toxin receptor from <i>Manduca sexta</i> , expressed in mammalian and insect cell cultures." APPLIED AND ENVIRONMENTAL MICROBIOLOGY, vol. 63, no. 9, 1997, pages 3419-3425, XP002164761 ISSN: 0099-2240 cited in the application the whole document	1,2,4, 9-15
X	KEETON TIMOTHY P ET AL: "Effects of midgut-protein-preparative and ligand binding procedures on the toxin binding characteristics of BT-R1, a common high-affinity receptor in <i>Manduca sexta</i> for CryIA <i>Bacillus thuringiensis</i> toxins." APPLIED AND ENVIRONMENTAL MICROBIOLOGY, vol. 64, no. 6, June 1998 (1998-06), pages 2158-2165, XP002164762 ISSN: 0099-2240 cited in the application the whole document	1,2,4, 9-15
A	GILL SARJEET S ET AL: "Identification, isolation, and cloning of a <i>Bacillus thuringiensis</i> CryIAC toxin-binding protein from the midgut of the lepidopteran insect <i>Heliothis virescens</i> ." JOURNAL OF BIOLOGICAL CHEMISTRY, vol. 270, no. 45, 1995, pages 27277-27282, XP002164763 ISSN: 0021-9258 cited in the application the whole document	

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# INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 00/31674

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	LEE MI KYONG ET AL: "Aminopeptidase N purified from gypsy moth brush border membrane vesicles is a specific receptor for Bacillus thuringiensis CryIAc toxin." APPLIED AND ENVIRONMENTAL MICROBIOLOGY, vol. 62, no. 8, 1996, pages 2845-2849, XP002164764 ISSN: 0099-2240 cited in the application the whole document	
E	WO 01 34807 A (CANDAS MEHMET ;BULLA LEE A JR (US)) 17 May 2001 (2001-05-17) claims 1-28; figures 1,2	1-15
A	GARCZYNSKI S F ET AL: "IDENTIFICATION OF PUTATIVE INSECT BRUSH BORDER MEMBRANE-BINDING MOLECULES SPECIFIC TO BACILLUS-THURINGIENSIS DELTA ENDOTOXIN BY PROTEIN BLOT ANALYSIS" APPLIED AND ENVIRONMENTAL MICROBIOLOGY, vol. 57, no. 10, 1991, pages 2816-2820, XP000992668 ISSN: 0099-2240 the whole document	
A	OLTEAN DANIELA I ET AL: "Partial purification and characterization of Bacillus thuringiensis CryIA toxin receptor A from Heliothis virescens and cloning of the corresponding cDNA." APPLIED AND ENVIRONMENTAL MICROBIOLOGY, vol. 65, no. 11, November 1999 (1999-11), pages 4760-4766, XP002169506 ISSN: 0099-2240 the whole document	

# INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US 00/31674

## Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:  
because they relate to subject matter not required to be searched by this Authority, namely:
2. ☐ Claims Nos.:  
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3. ☐ Claims Nos.:  
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

## Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

see additional sheet

1. ☒ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

### Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☒ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ASA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: (1-25)-partially

An isolated nucleic acid molecule having the nucleotide sequence encoding a Bt toxin receptor, selected from *Ostrinia nubilalis* respectively SEQ ID No. 1; said nucleic acid, wherein said toxin is CryIA/CryIA(b); an isolated polypeptide having the amino acid sequence set forth in SEQ ID NO. 2; a fusion polypeptide comprising said polypeptide, an expression cassette comprising a nucleotide sequence encoding said fusion polypeptide; an antibody preparation specific for said polypeptide; a vector comprising said expression cassette; a cell comprising said vector; a transformed cell of interest having stably incorporated within its genome said nucleotide sequence, SEQ ID No.1; a method for screening for ligands that bind said Bt toxin receptor comprising SEQ ID No. 2;

2. Claims: (1-25)-partially

Idem as invention 1 but limited to *Heliothis zea*, respectively SEQ ID Nos. 3 and 4;

3. Claims: (1-25)-partially

Idem as invention 1 but limited to *Spodoptera frugiperda*, respectively SEQ ID Nos. 5 and 6;



# INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 00/31674

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9612964 A	02-05-1996	US 5693491 A	02-12-1997
		AU 711066 B	07-10-1999
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		CA 2200427 A	02-05-1996
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		JP 10508198 T	18-08-1998
		NZ 296265 A	28-05-1999
		US 6007981 A	28-12-1999
		ZA 9508851 A	11-06-1996
WO 0134807 A	17-05-2001	NONE	

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